



# “Live Cadaver”

Is there spare space for cadavers  
in surgical training?

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Emad Aboud, M.D

Arkansas Neuroscience Institute

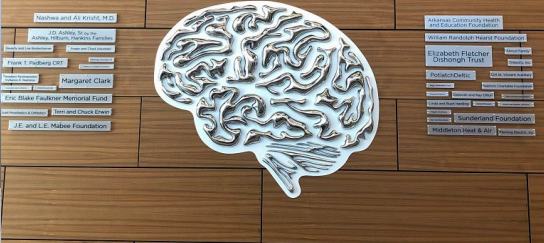


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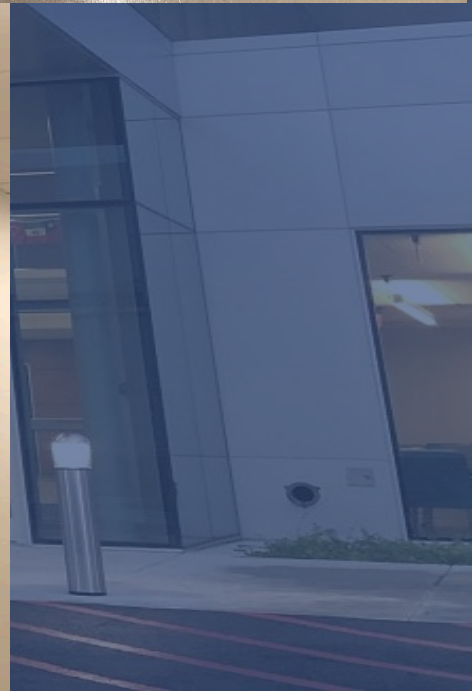
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### ARKANSAS NEUROSCIENCE INSTITUTE RESEARCH & EDUCATION CENTER



# OSSAMA AL-MEFTY MICRONEUROSURGERY LABORATORY



**Nothing to disclose**

See one, Do one, Teach one.

Is not a valid strategy anymore.

There is a need an adjacent methods  
in addition to the OR experience to  
teach residents how to do surgery.

## **Available Training Models**

- Virtual Reality/ computer model
- Physical models/high fidelity simulators
  - Live animals
  - Human cadavers

## Available Training Models

- Virtual Reality/ computer model
- Physical models/high fidelity simulators
  - Live animals
  - Human cadavers

# - Virtual Reality/ computer model



TECHNIQUE ASSESSMENT

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Cristian J. Luciano, PhD†§  
Daniel P. Bailey, PhD‡  
Abdussalam Elsenousi, MD\*  
Ben Z. Rothberg, MD|  
Antonio Bernardo, MD#  
P. Pat Banerjee, PhD||§  
Fady T. Charbel, MD\*

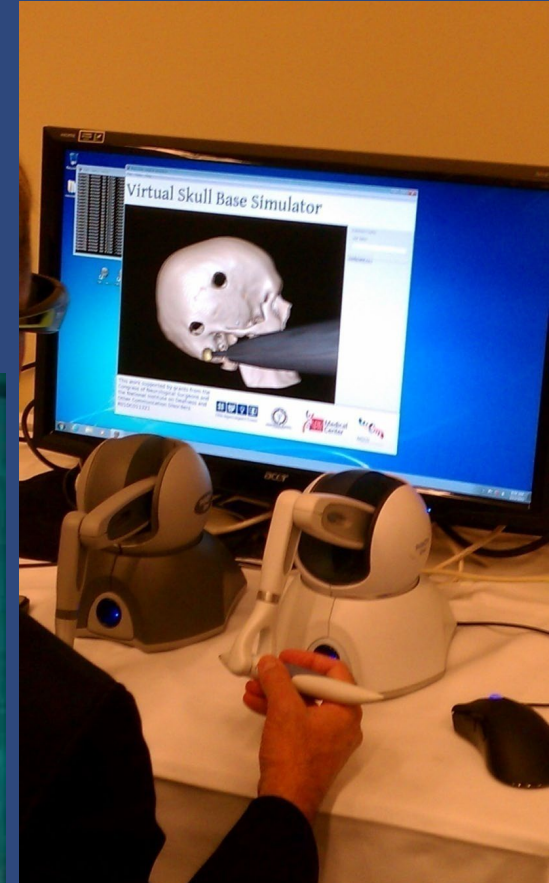
**Virtual Reality Cerebral Aneurysm Clipping Simulation With Real-Time Haptic Feedback**

**BACKGROUND:** With the decrease in the number of cerebral aneurysms treated surgically and the increase of complexity of those treated surgically, there is a need for simulation-based tools to teach future neurosurgeons the operative techniques of aneurysm clipping.

**OBJECTIVE:** To develop and evaluate the usefulness of a new haptic-based virtual reality simulator in the training of neurosurgical residents.

**METHODS:** A real-time sensory haptic feedback virtual reality aneurysm clipping simulator was developed using the ImmersiveTouch platform. A prototype middle cerebral artery aneurysm simulation was created from a computed tomographic angiogram. Aneurysm and vessel volume deformation and haptic feedback are provided in a 3-dimensional immersive virtual reality environment. Intraoperative aneurysm rupture was also simulated. Seventeen neurosurgery residents from 3 residency programs tested the simulator and provided feedback on its usefulness and resemblance to real aneurysm clipping surgery.

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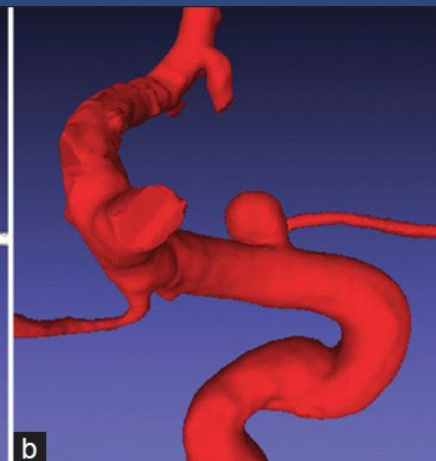
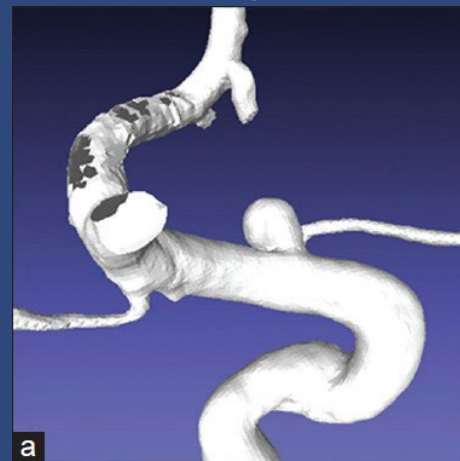


## **Available Training Models**

- Virtual Reality/ computer model
- Physical models/high fidelity simulators
  - Live animals
  - Human cadavers



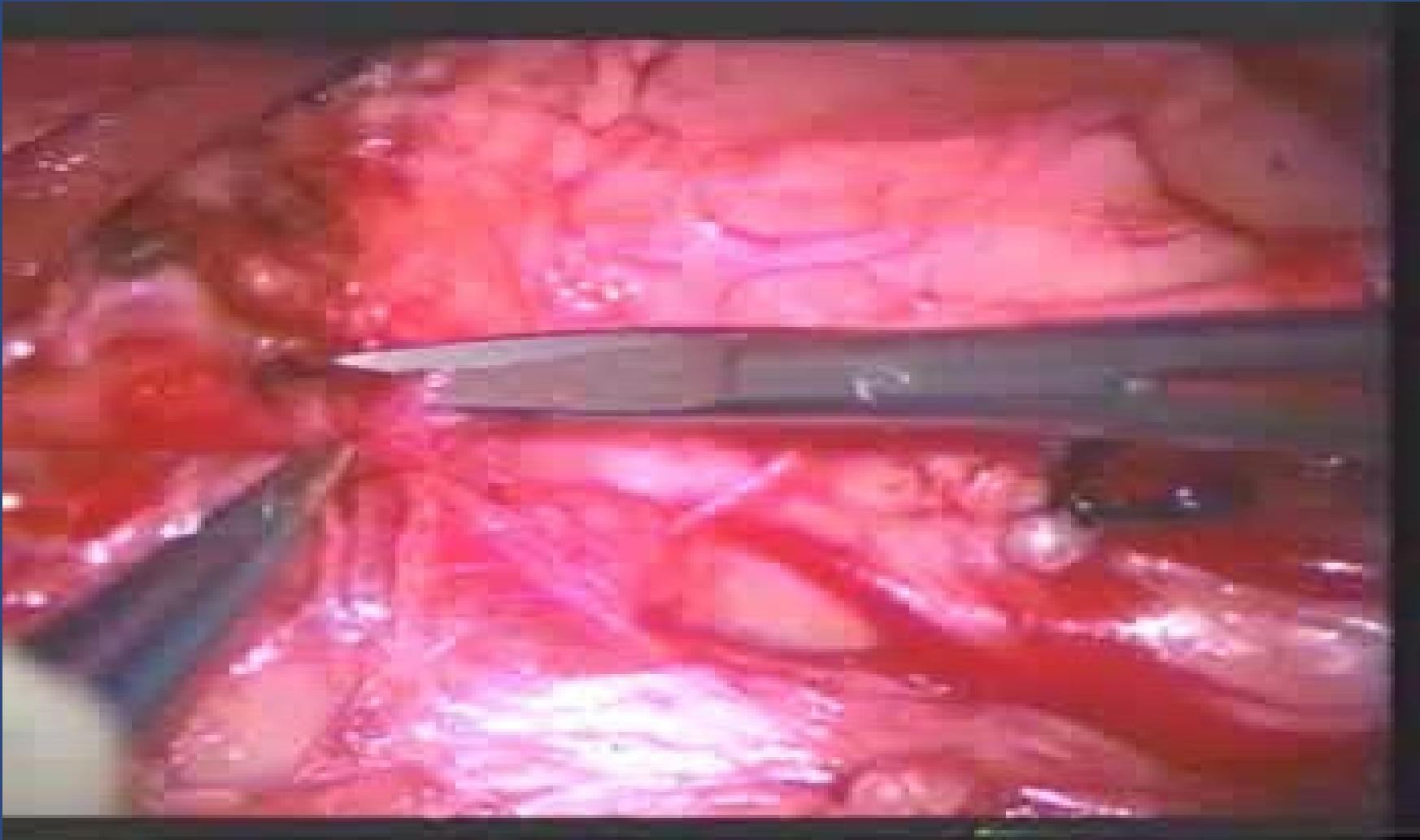
# - Physical models/high fidelity simulators



## **Available Training Models**

- Virtual Reality/ computer model
- Physical models/high fidelity simulators
  - **Live animals**
  - Human cadavers

- Live animals



## **Available Training Models**

- Virtual Reality/ computer model
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- Human cadavers

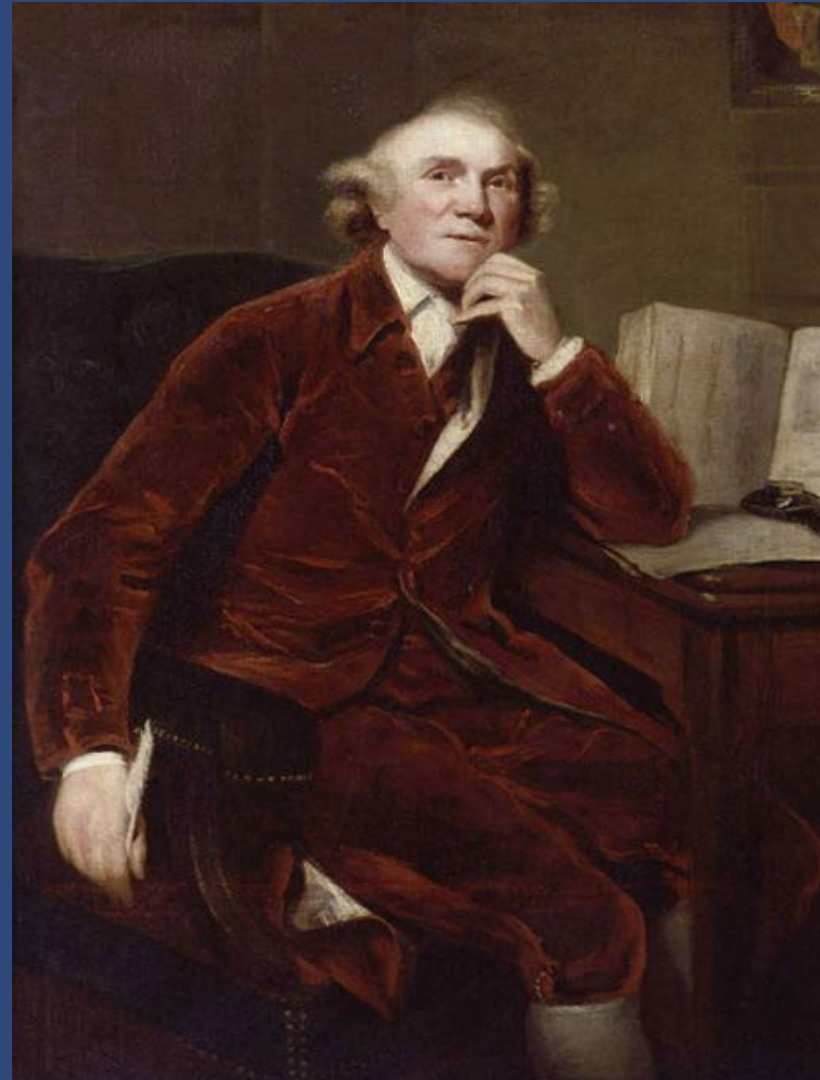


## **We Need a Training Model**

conveys a sense of reality to replicate live surgery, by providing the anatomical characteristics, **life-like** presentation, and **ability to bleed**.

# John Hunter

18<sup>th</sup> Century



## The transition from hunterian ligation to intracranial aneurysm clips: a historical perspective

**NIKA V. POLEVAYA, B.S., M. YASHAR S. KALANI, PH.D., GARY K. STEINBERG, M.D., PH.D.,  
AND VICTOR C. K. TSE, M.D., PH.D.**

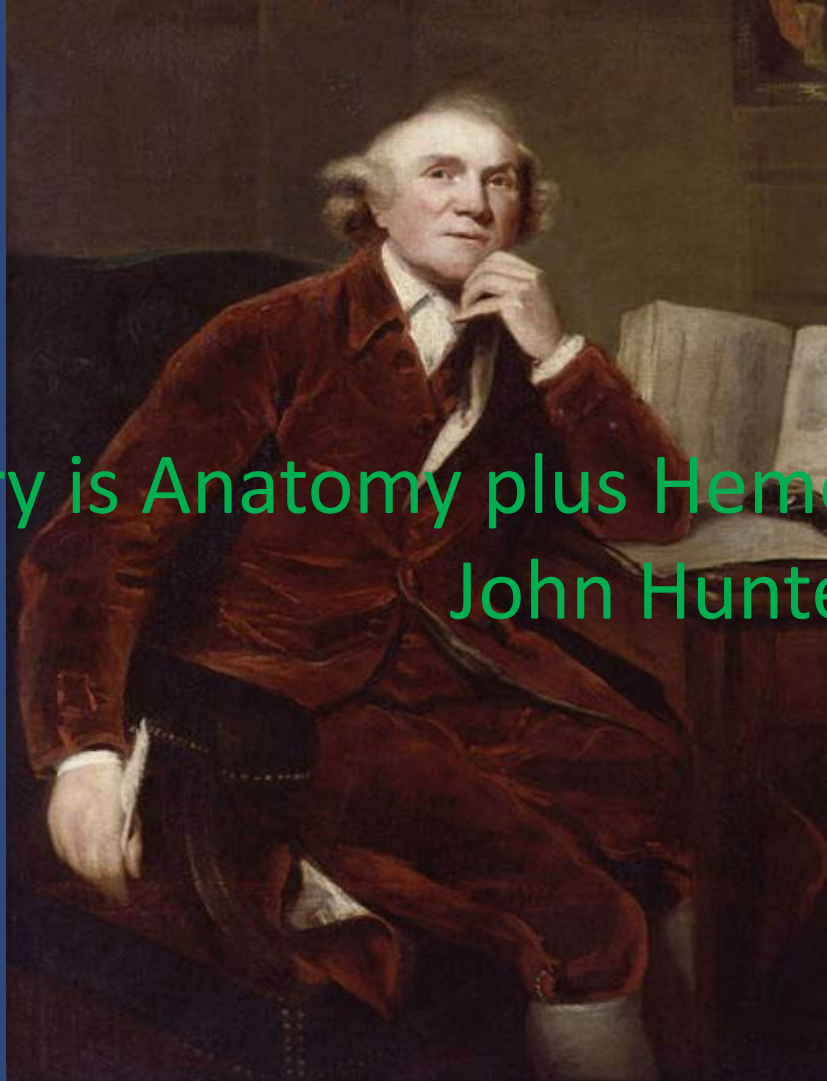
*Departments of Neurosurgery and Developmental Biology, Stanford University School of Medicine  
Stanford, California; and Howard Hughes Medical Institute, Chevy Chase, Maryland*

✓ The description of cerebral aneurysms dates back to antiquity. Little was known, however, about the pathological mechanisms of aneurysm formation and treatment options for this disease until 200 years ago. The modern era of aneurysm treatment began with the hunterian ligation of the proximal artery, followed by clip and coil occlusion. In this article, the authors describe the transition from conservative therapy to internal carotid artery (ICA) ligation and gradual occlusion of the ICA to the direct placement of clips on aneurysms. The driving forces and rationale behind each major advancement are summarized, and the authors attempt to predict what these innovations mean for the future of intracranial aneurysm management.

**KEY WORDS • intracranial aneurysm • clip occlusion • history of neurosurgery**

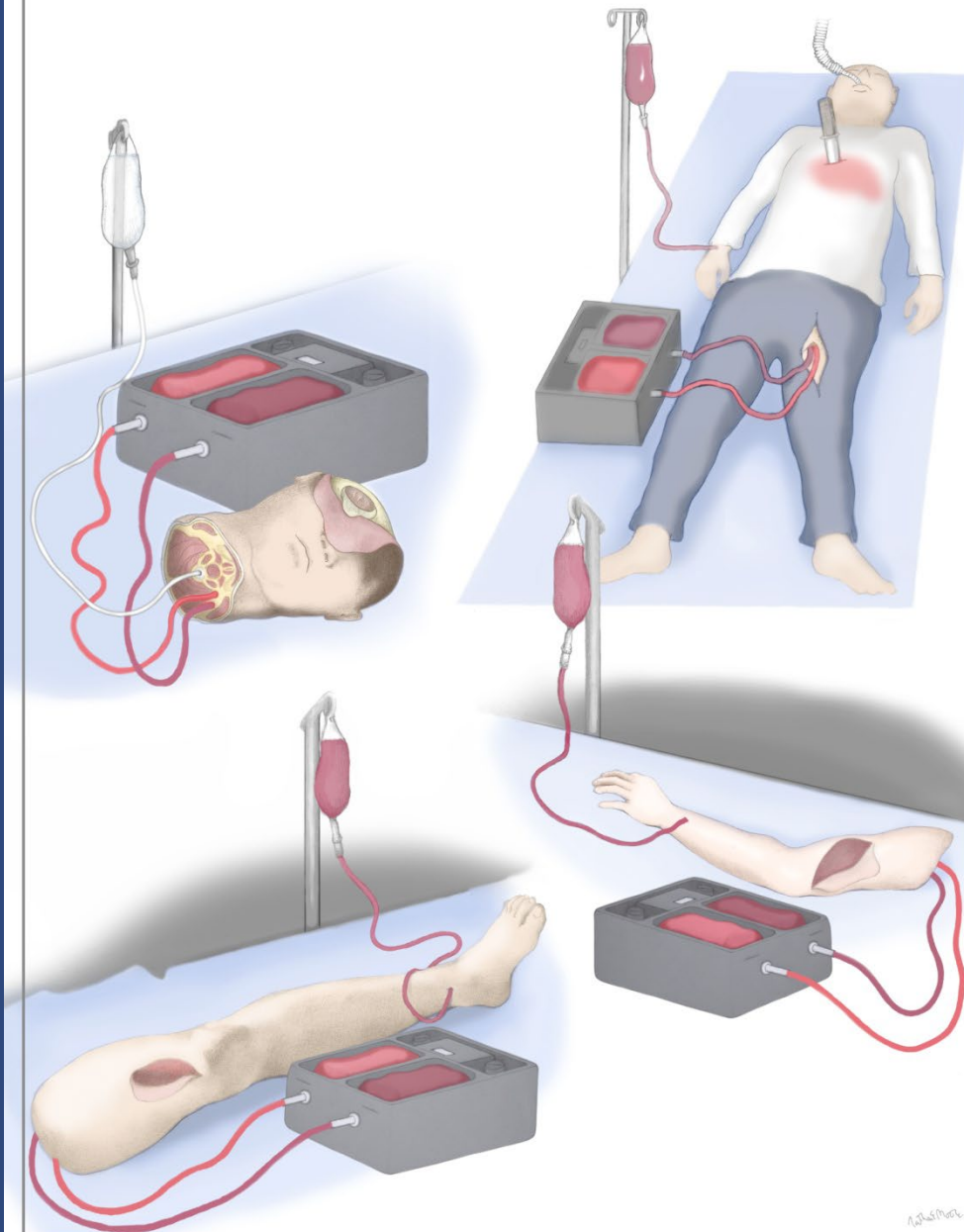


Surgery is Anatomy plus Hemostasis.  
John Hunter



# LIVE CADAVER MODEL

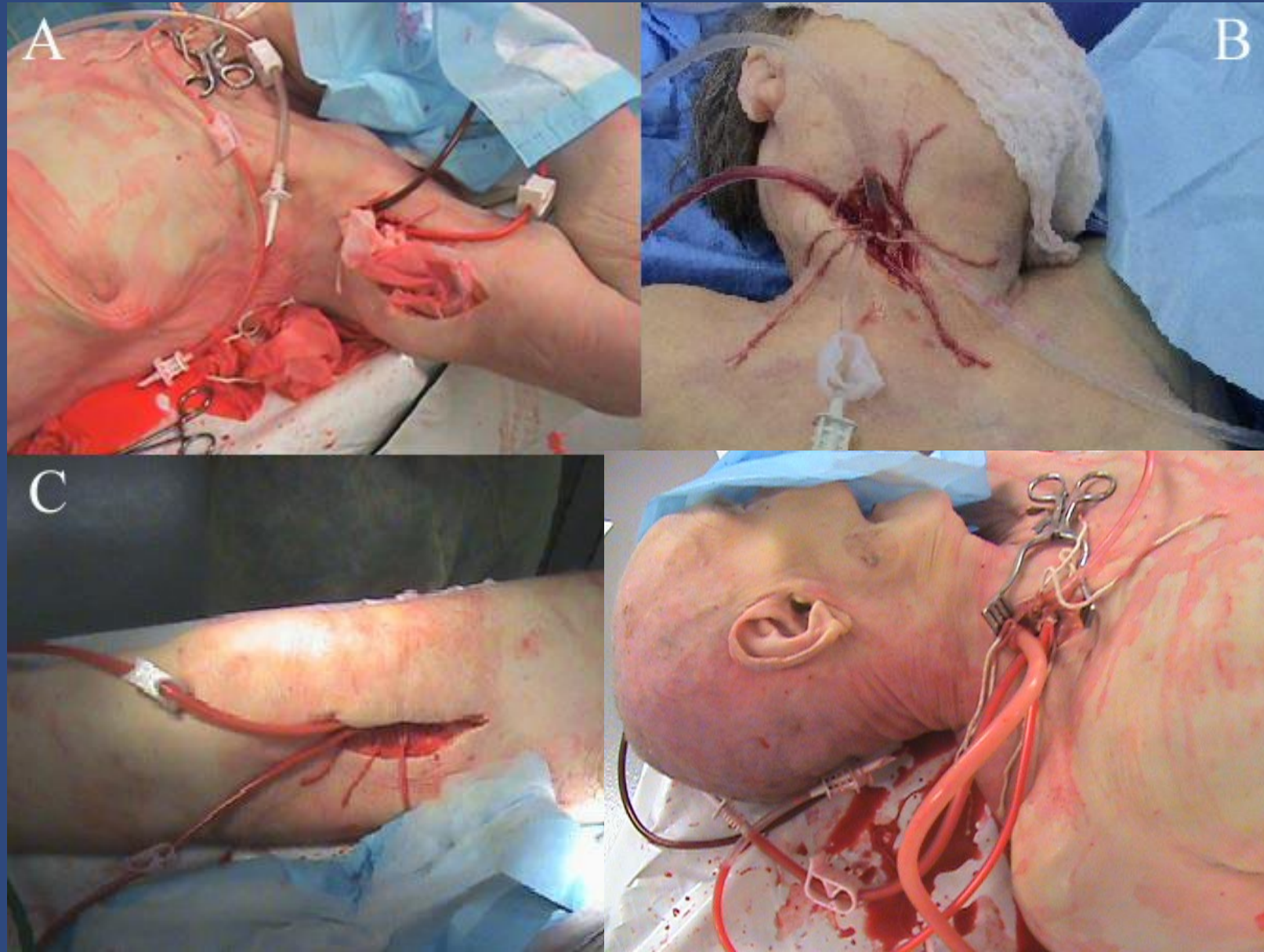
(ABOUD MODEL)





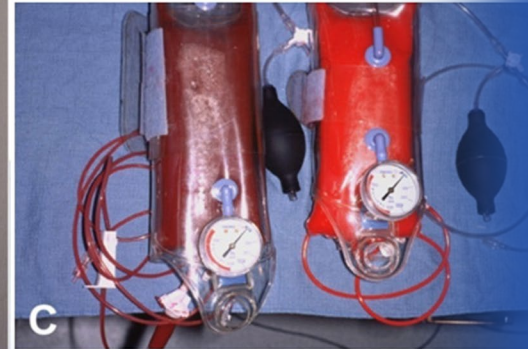
A Cadaver based model  
that combines the Real Human Anatomy  
with the  
life like condition of the living body

Canulate the vessels using any possible route  
We usually use the neck or the groin on one side



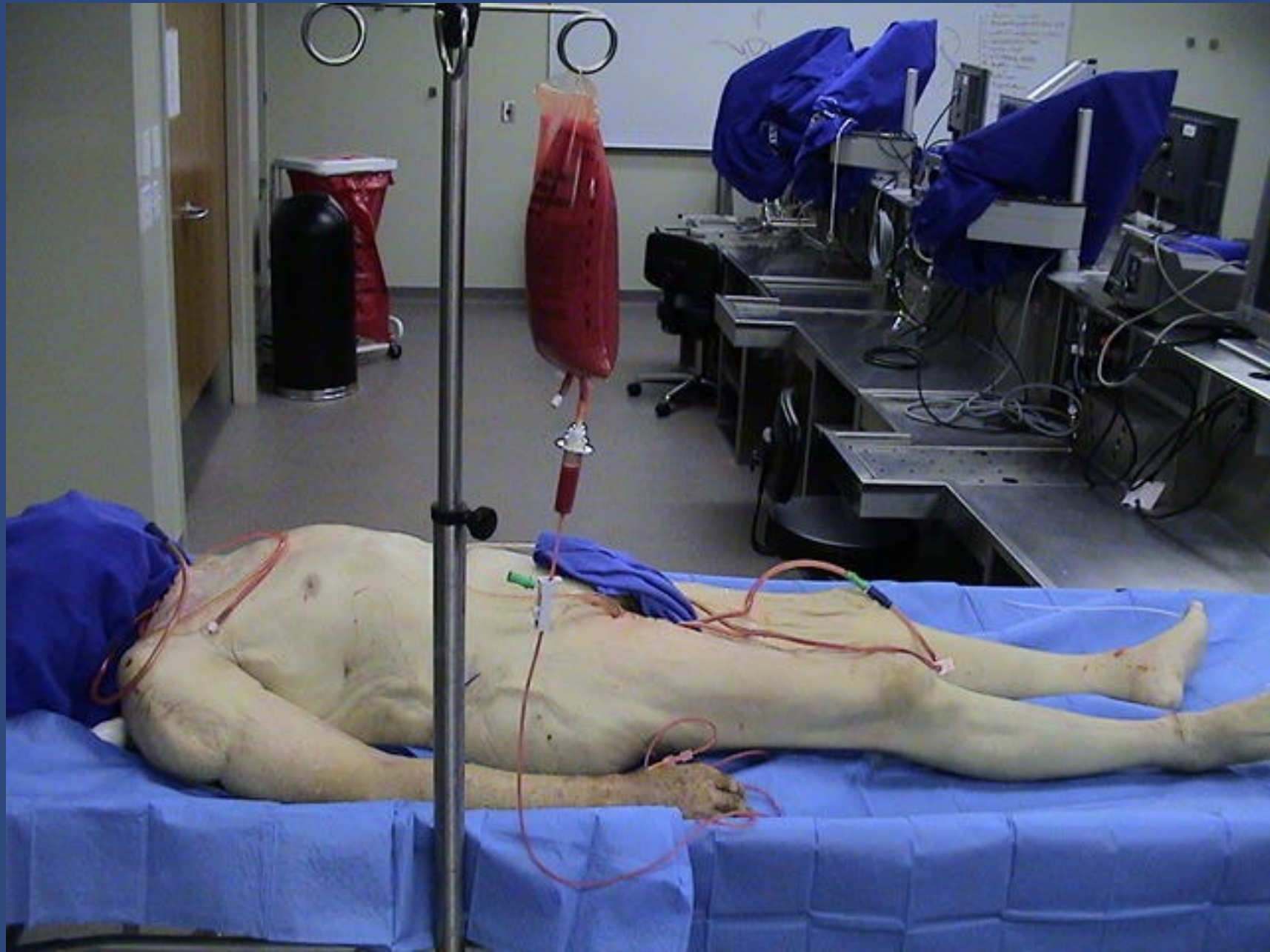


connect the cannulated vessels  
To artificial blood reservoirs and a  
machine that provides pulsating pressure

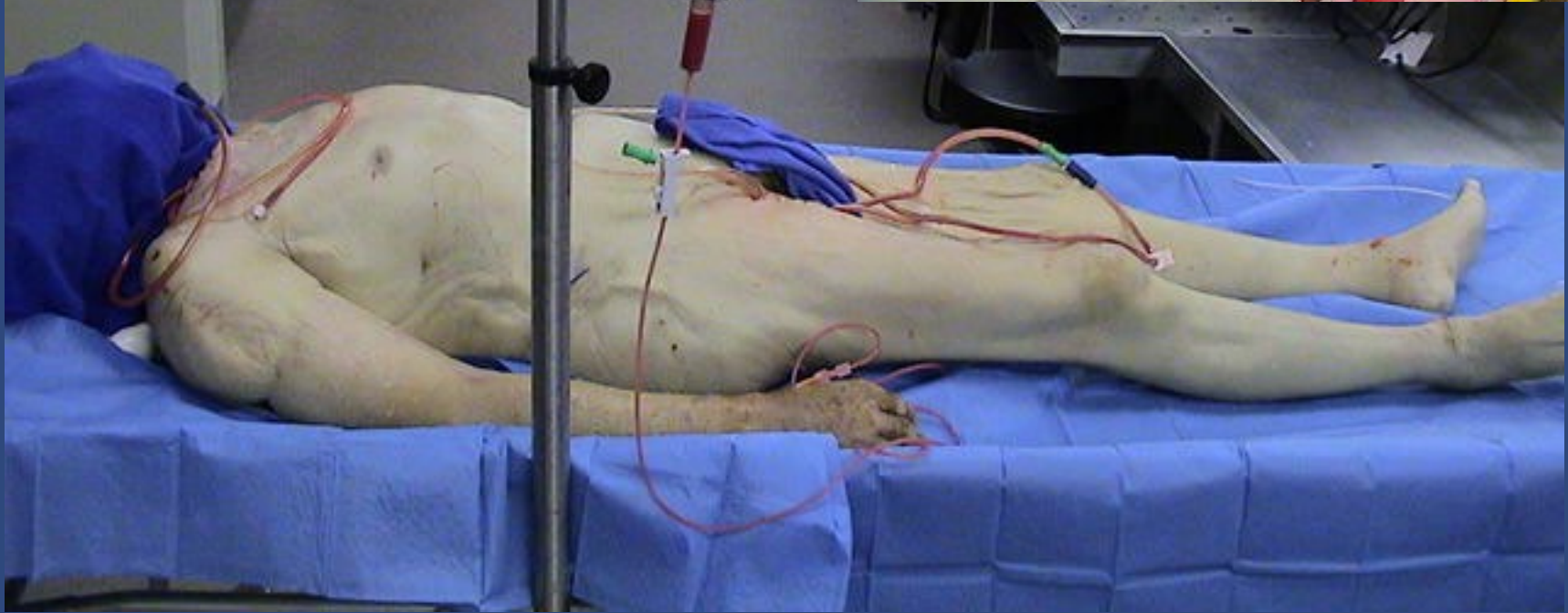


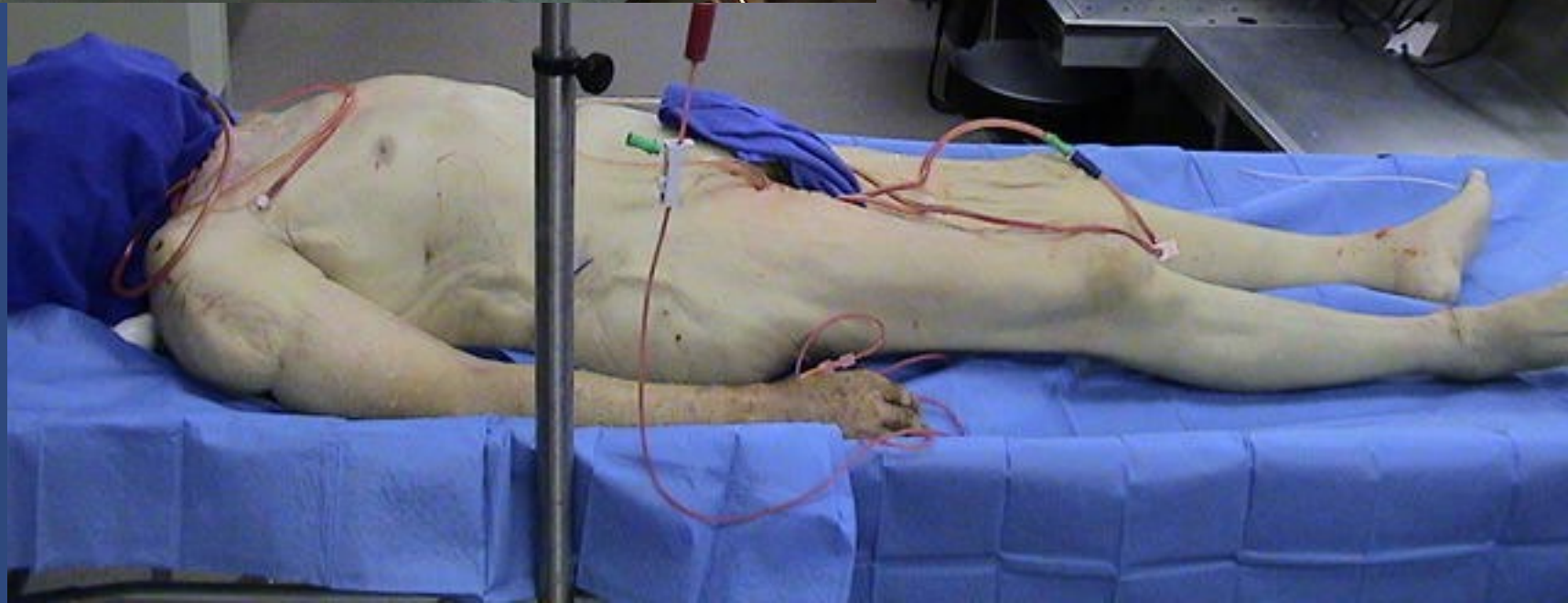
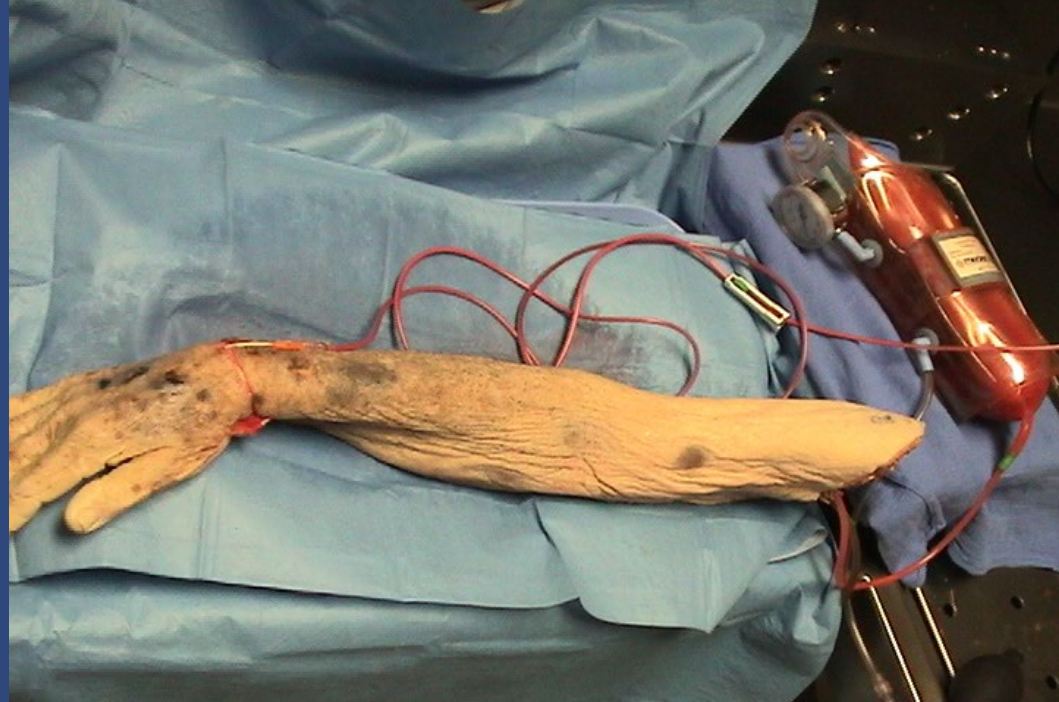
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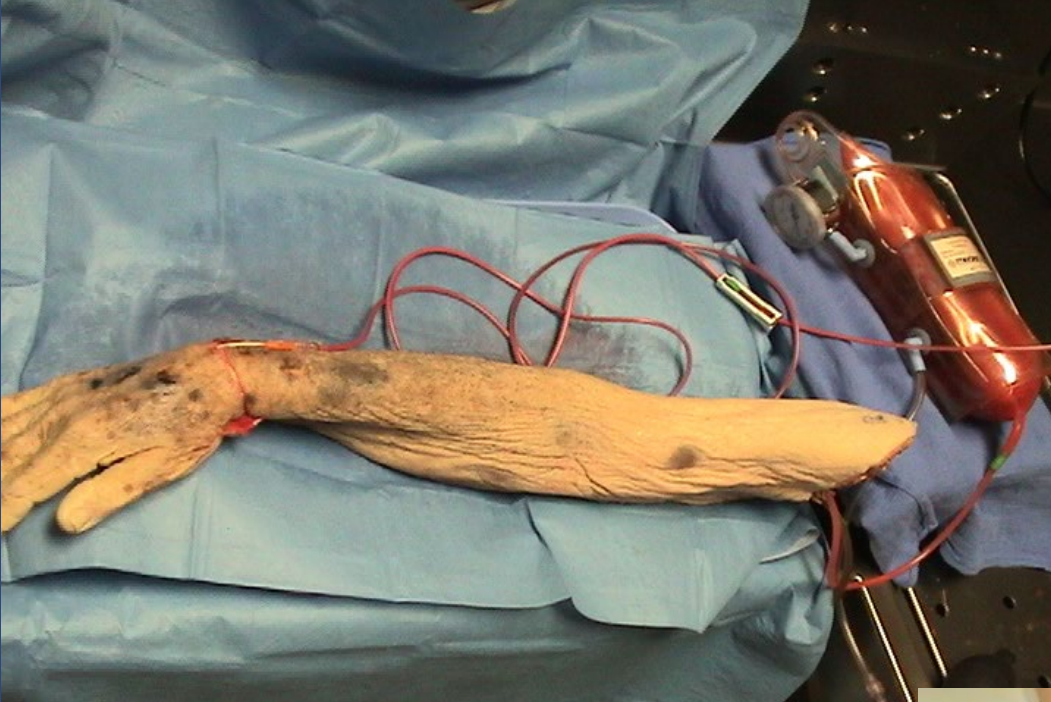


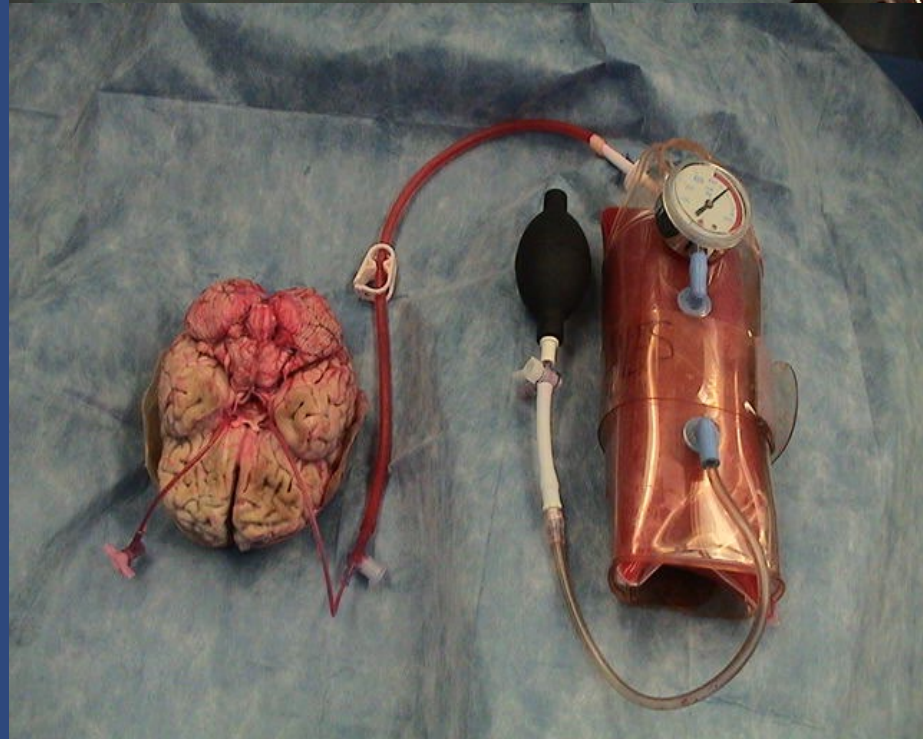


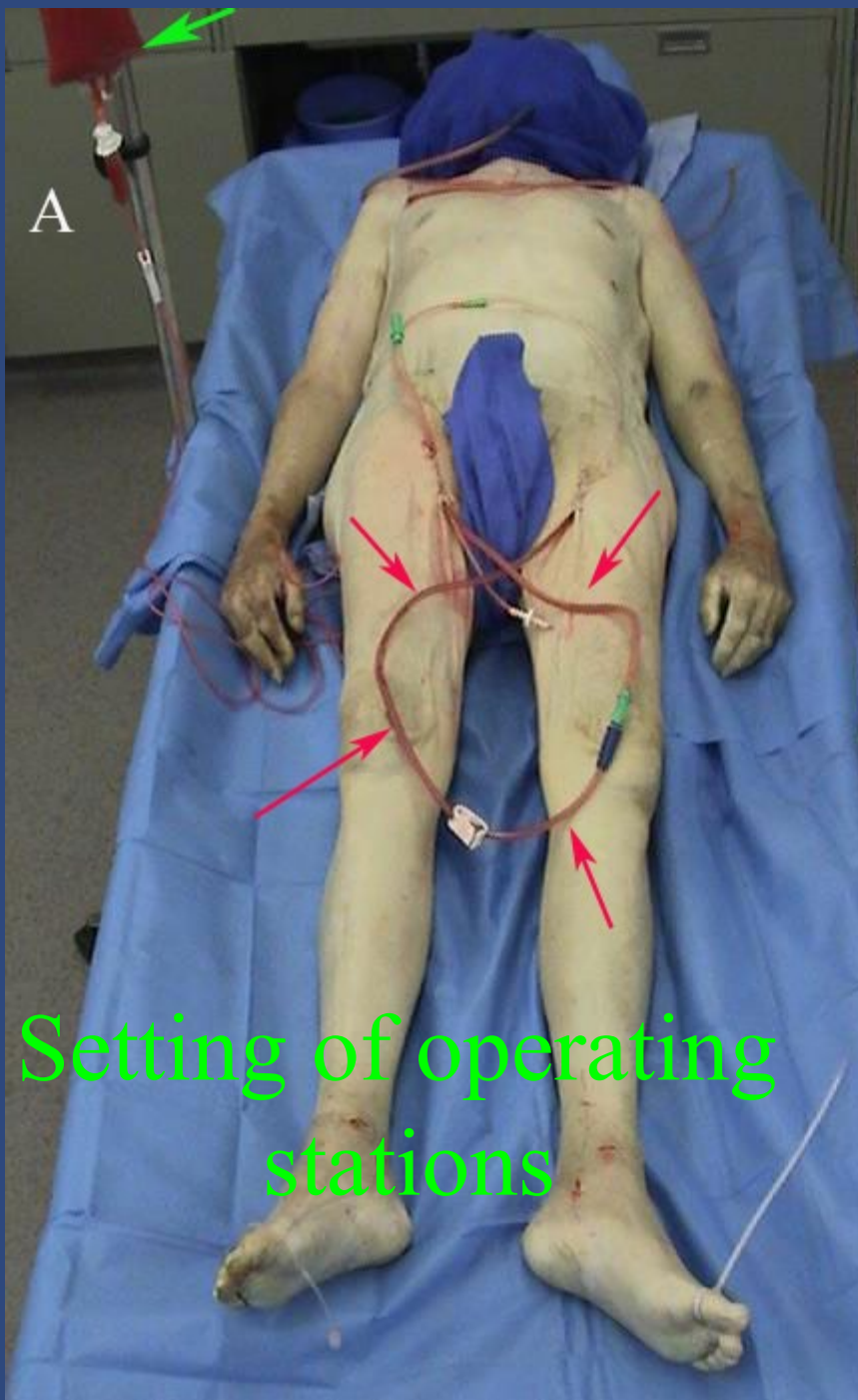


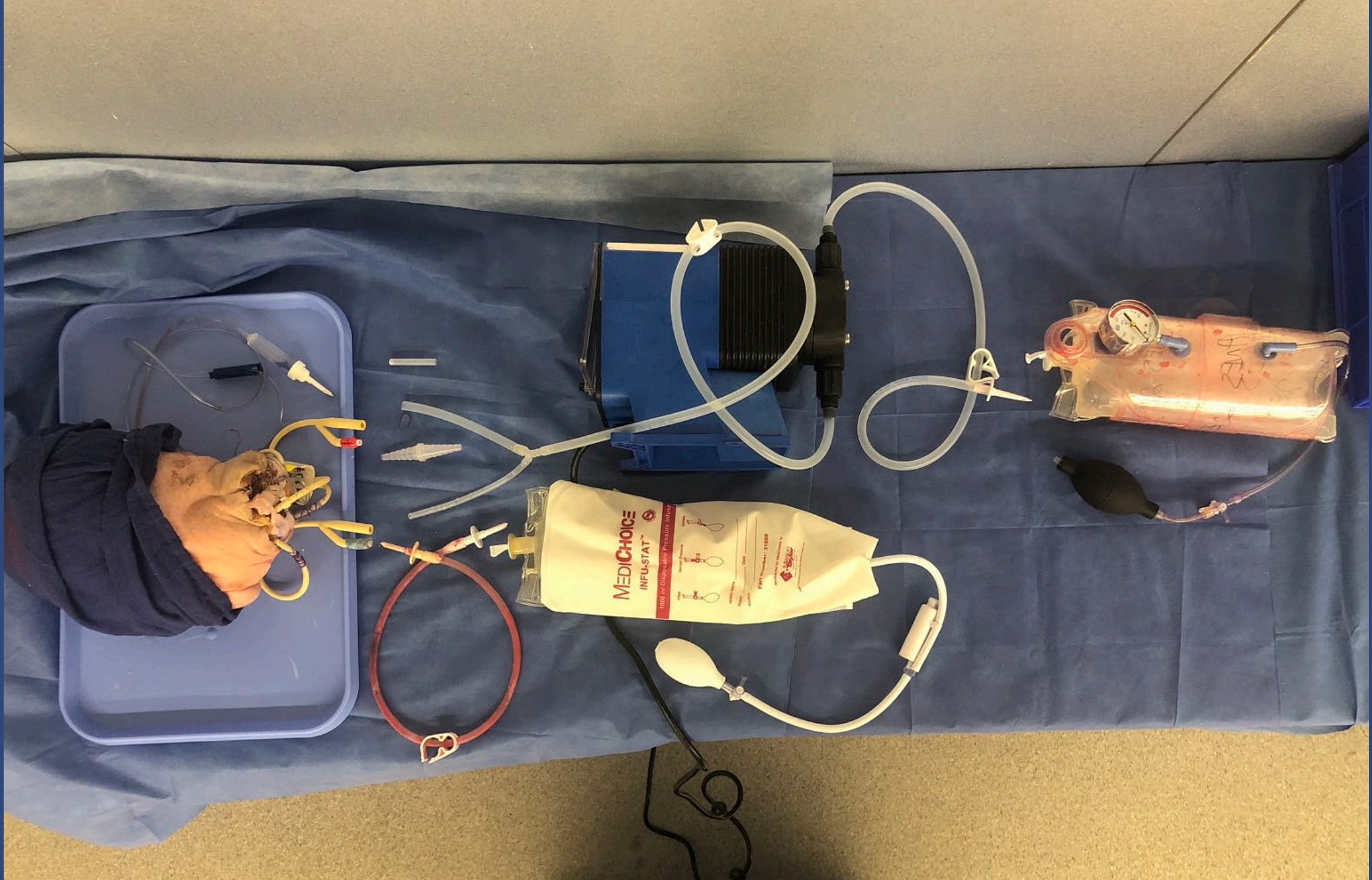


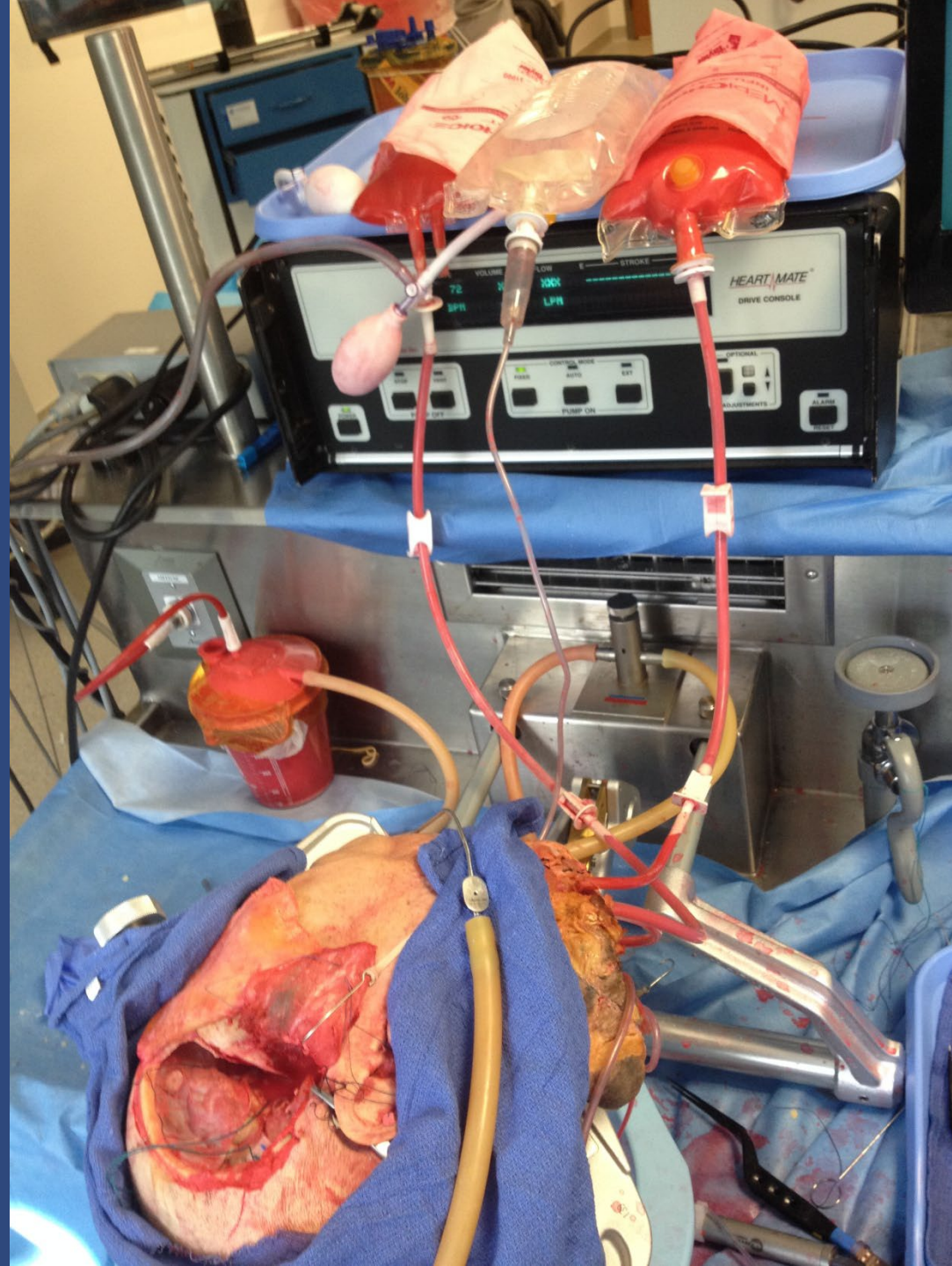






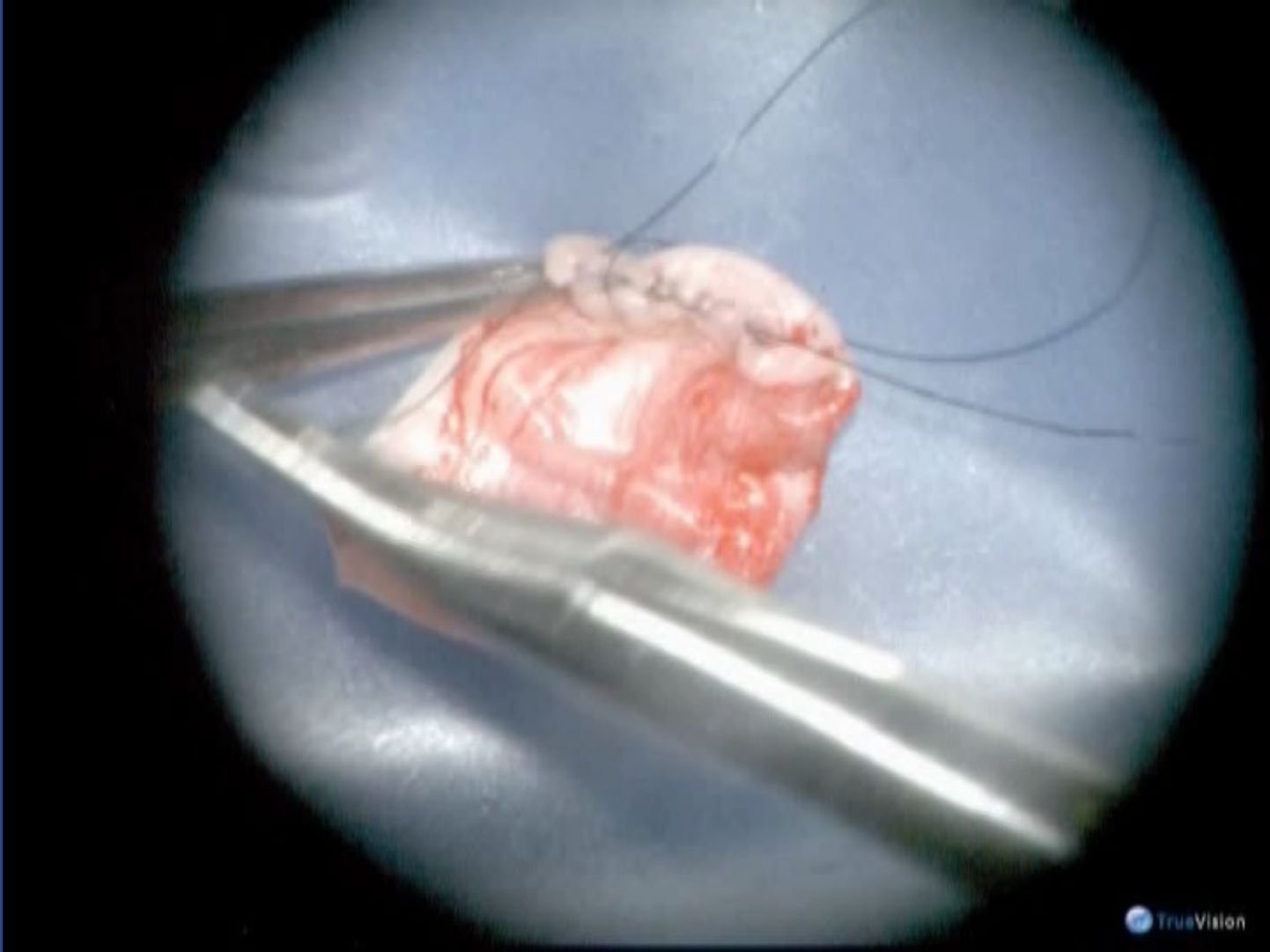


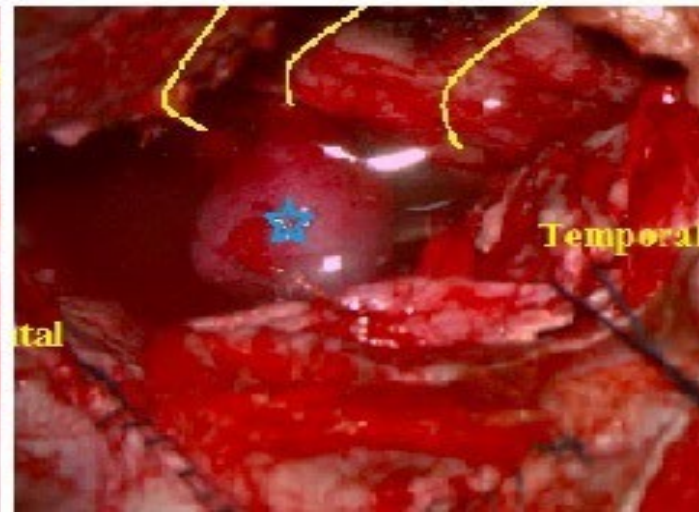
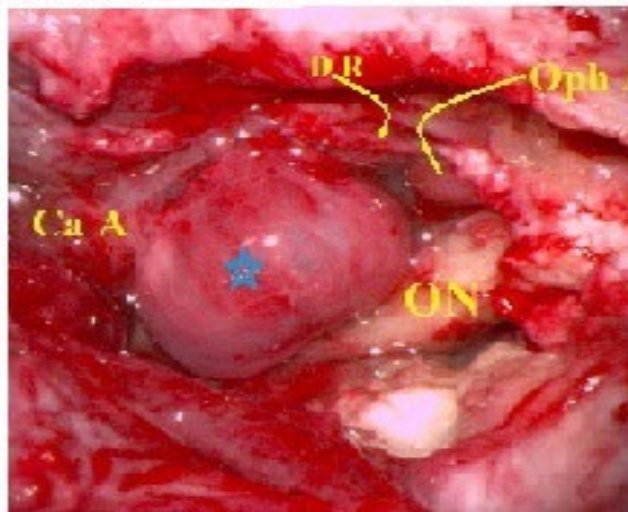
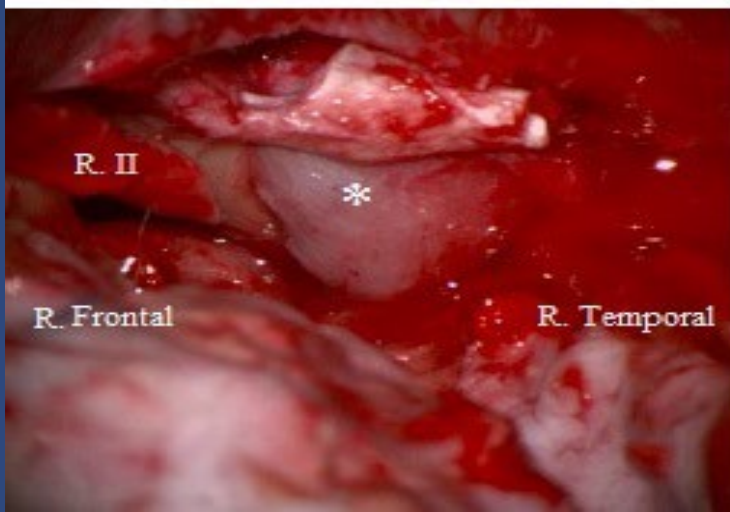
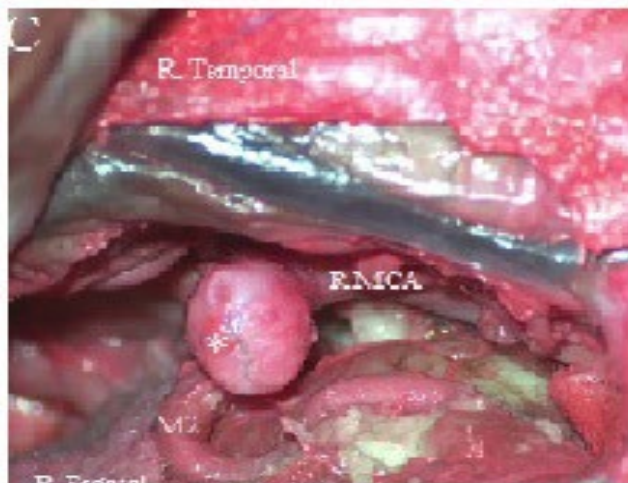
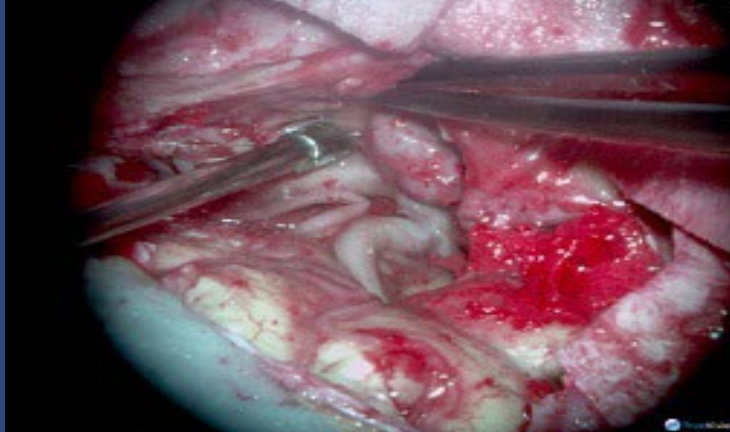


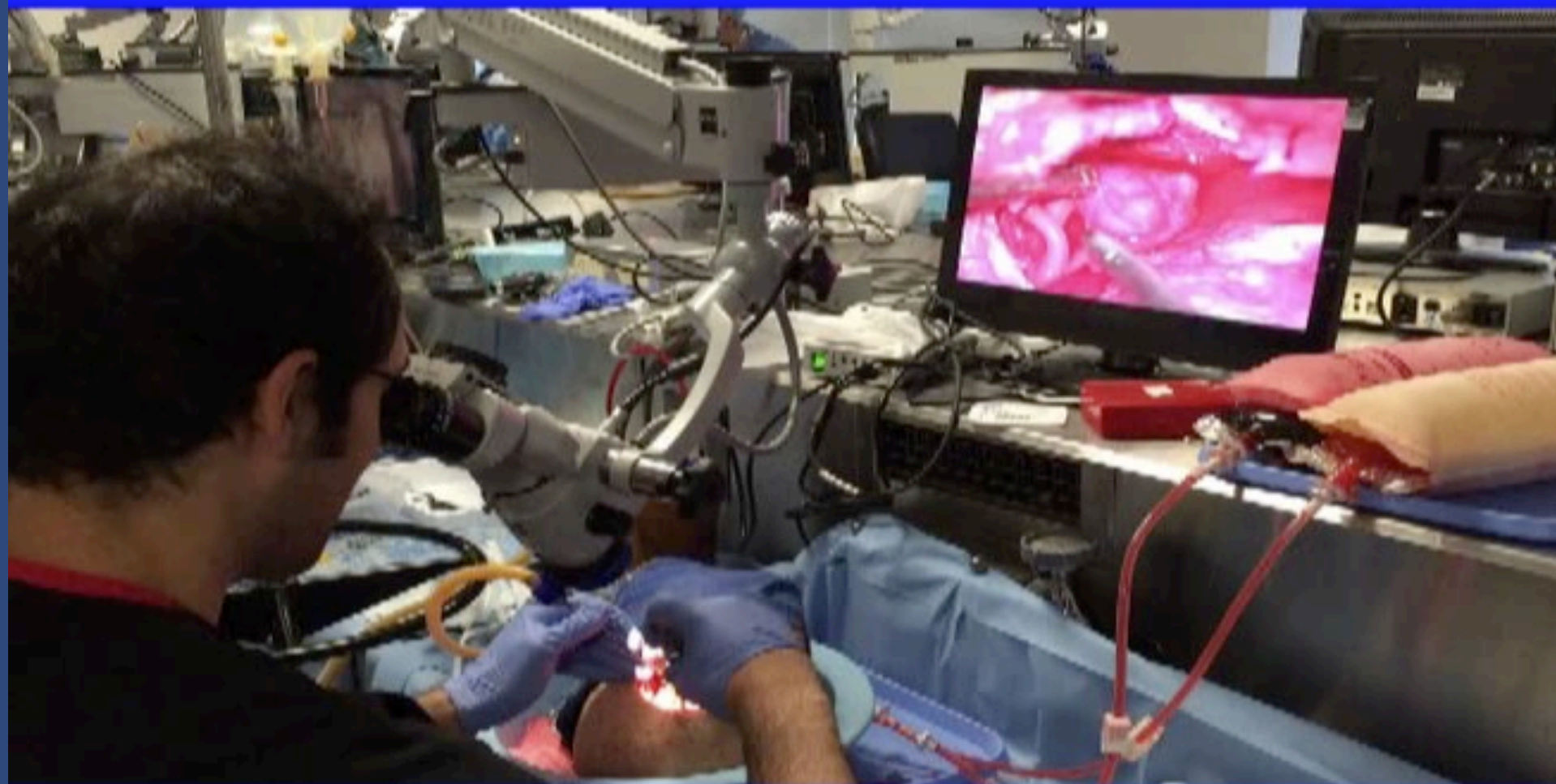




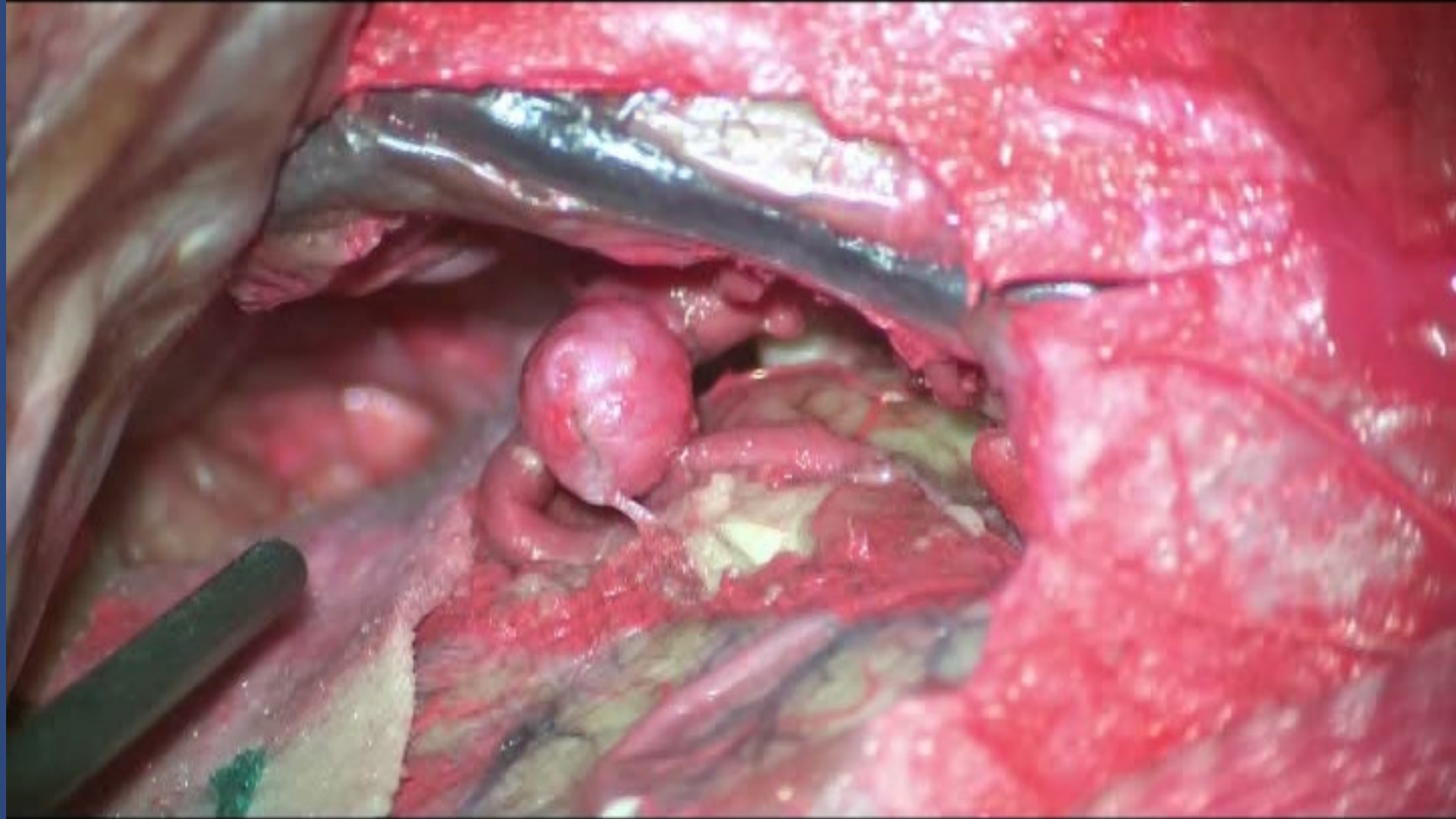


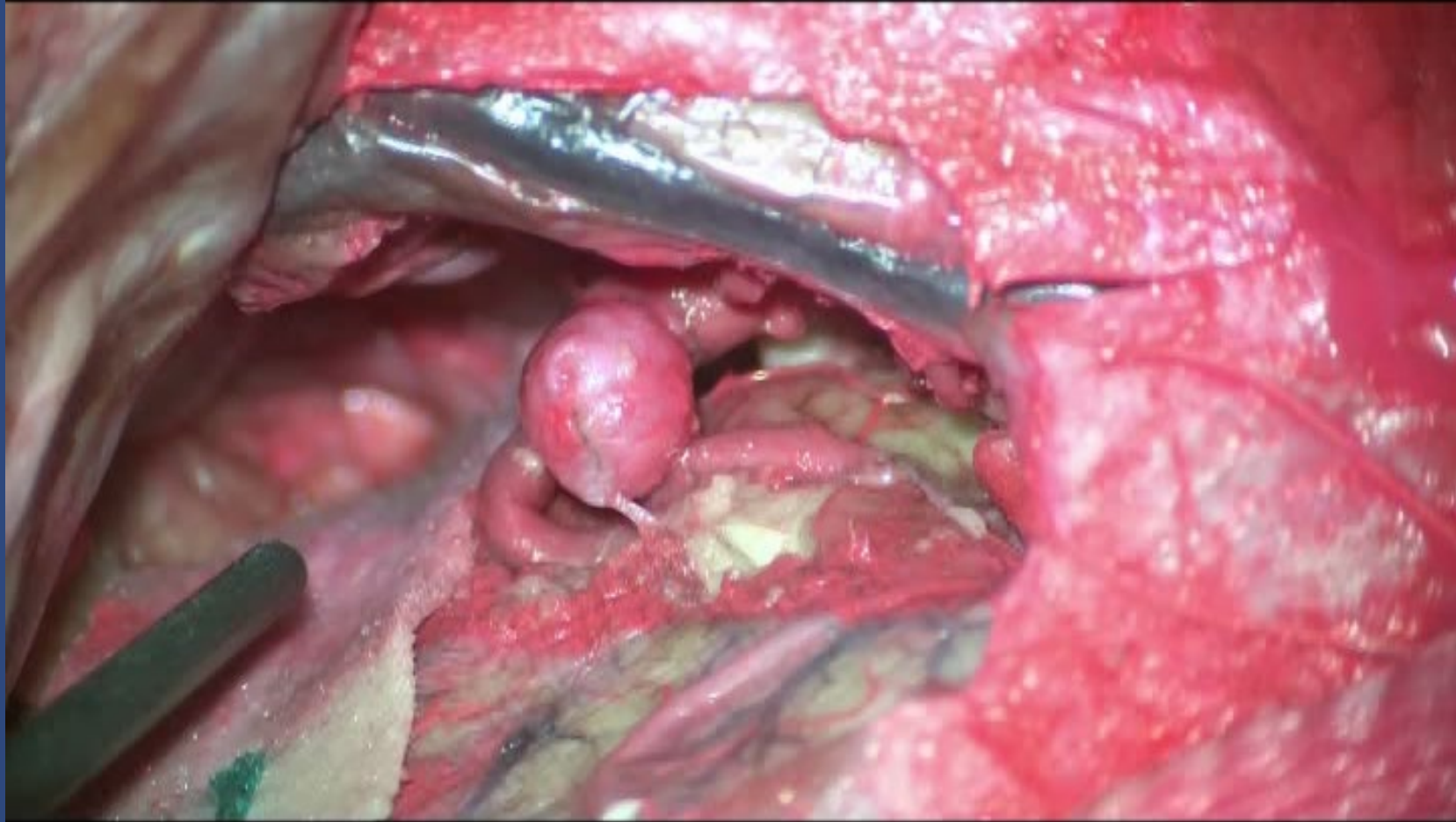












DAD ANGIO MODEL

SR. RAJESH K. S. (10/10/2020)

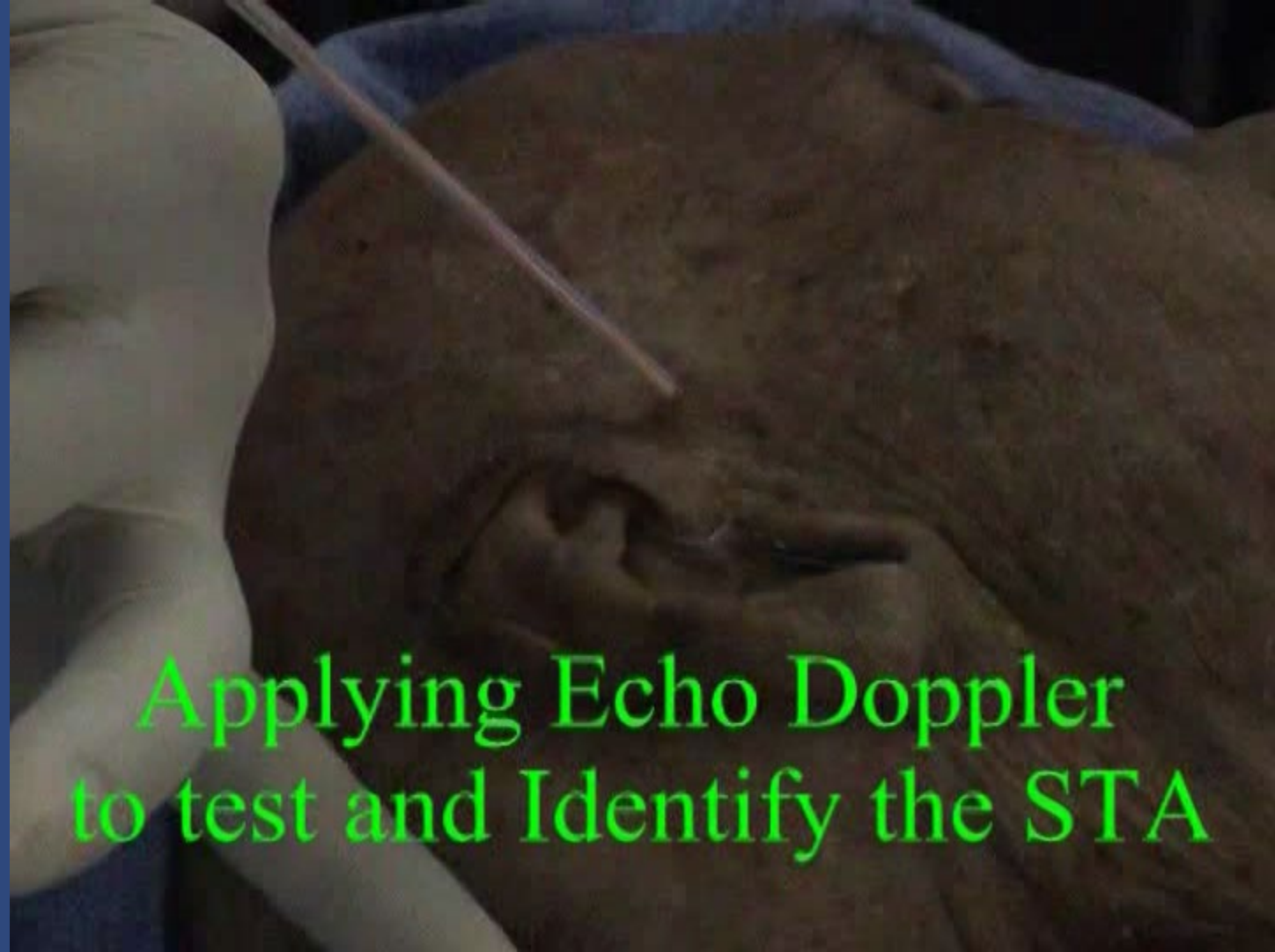
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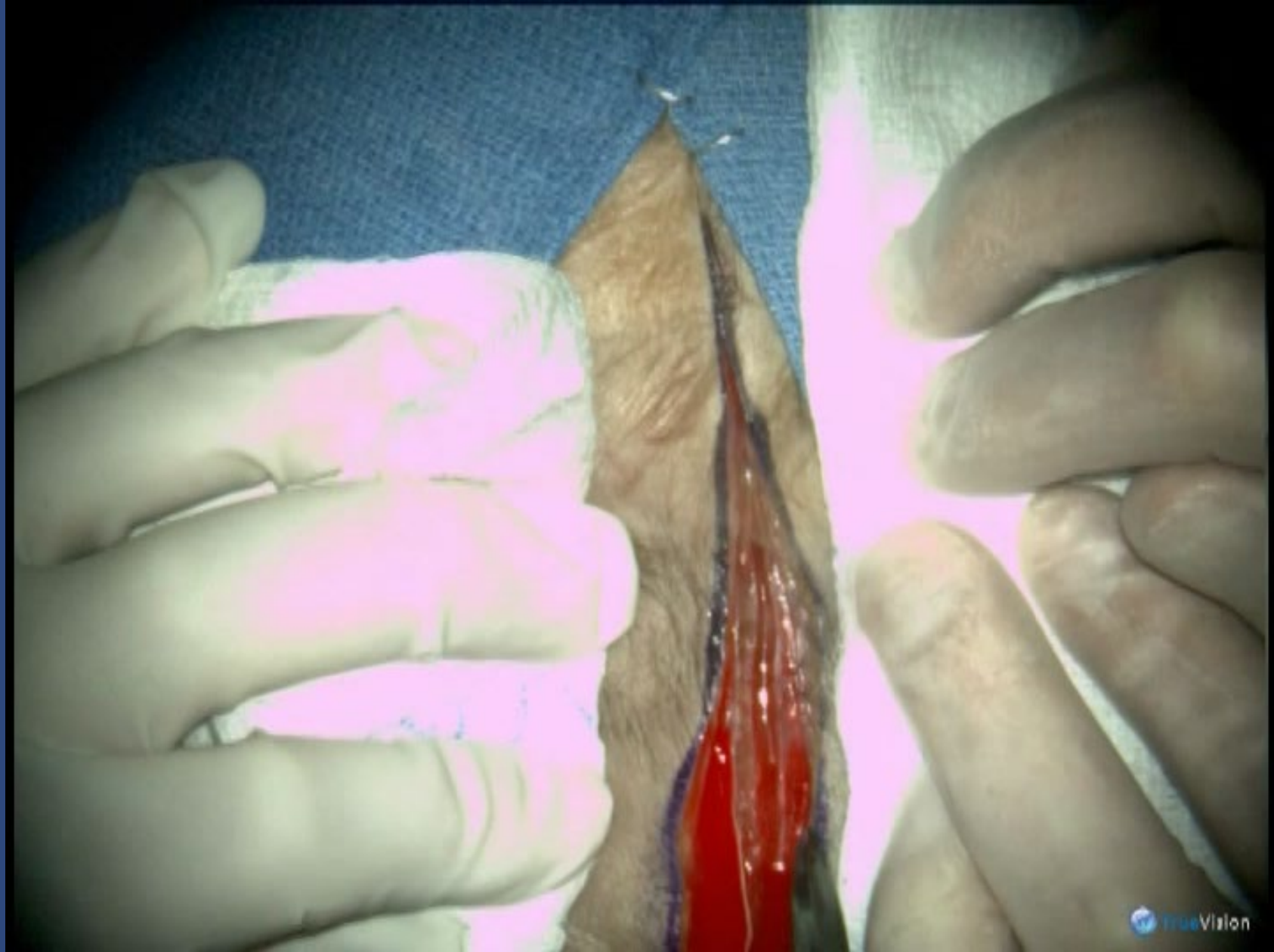
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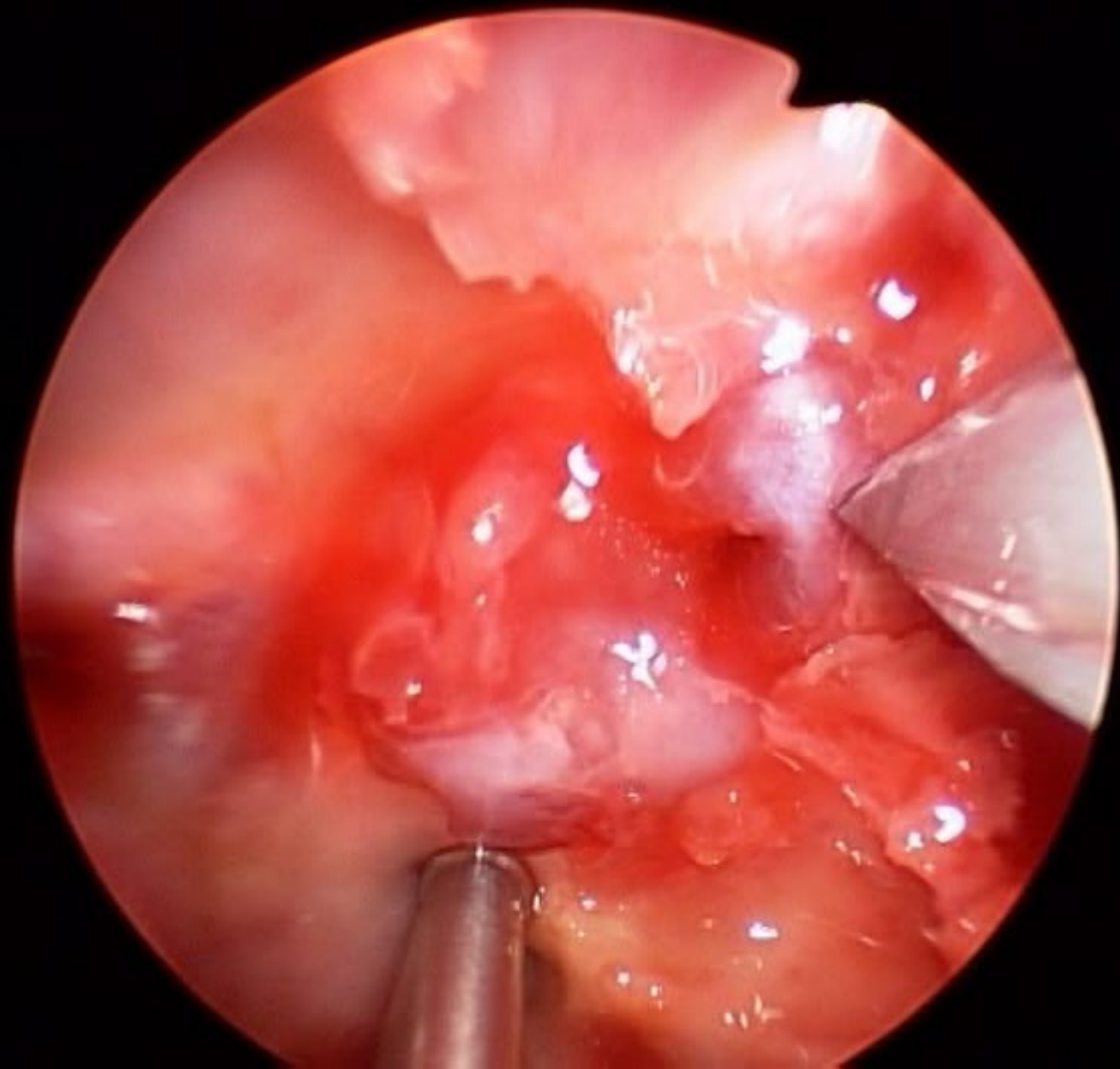
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Applying Echo Doppler  
to test and Identify the STA

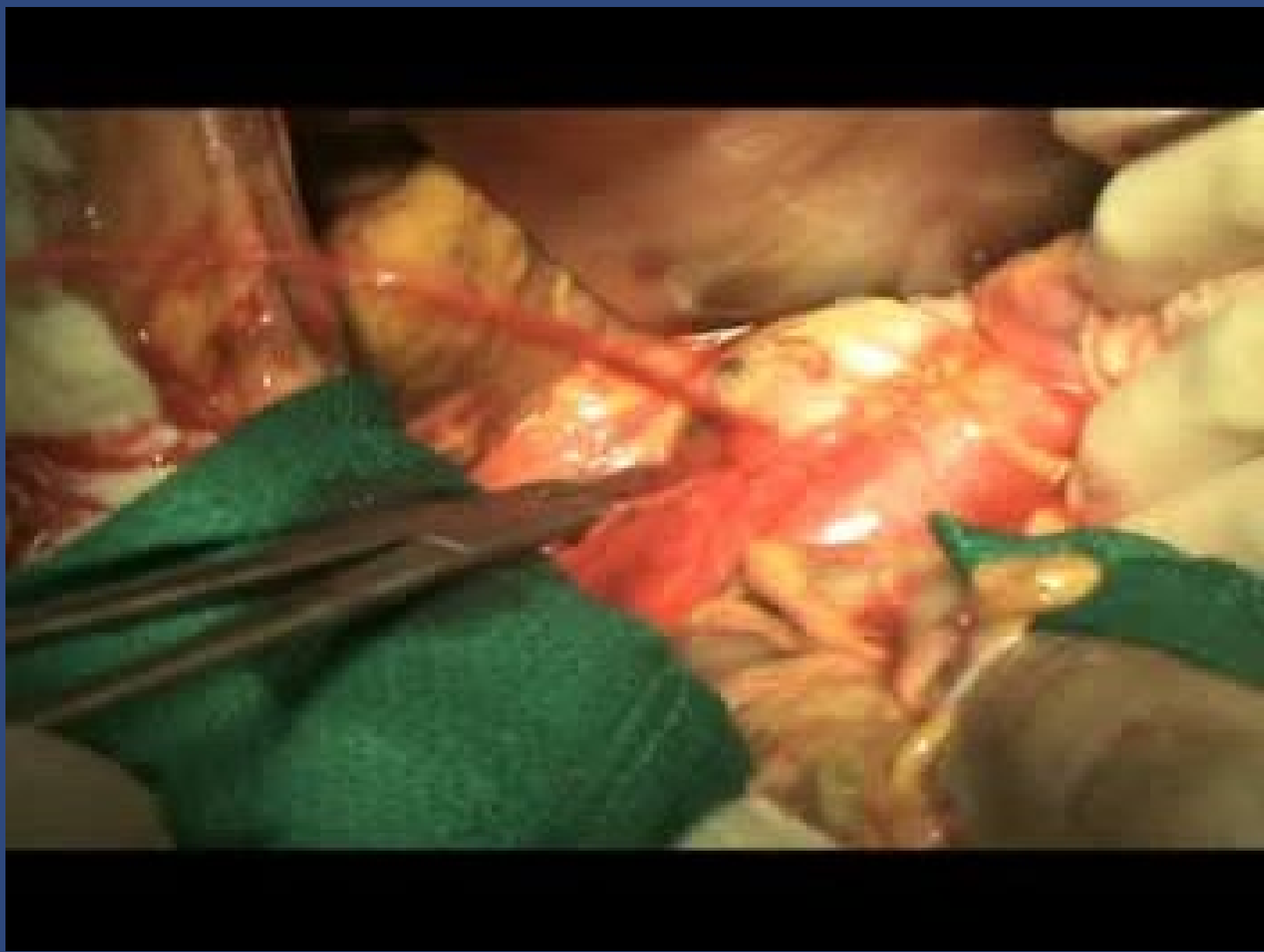






Skull Base Society  
4th annual meeting  
Memphis Feb 03

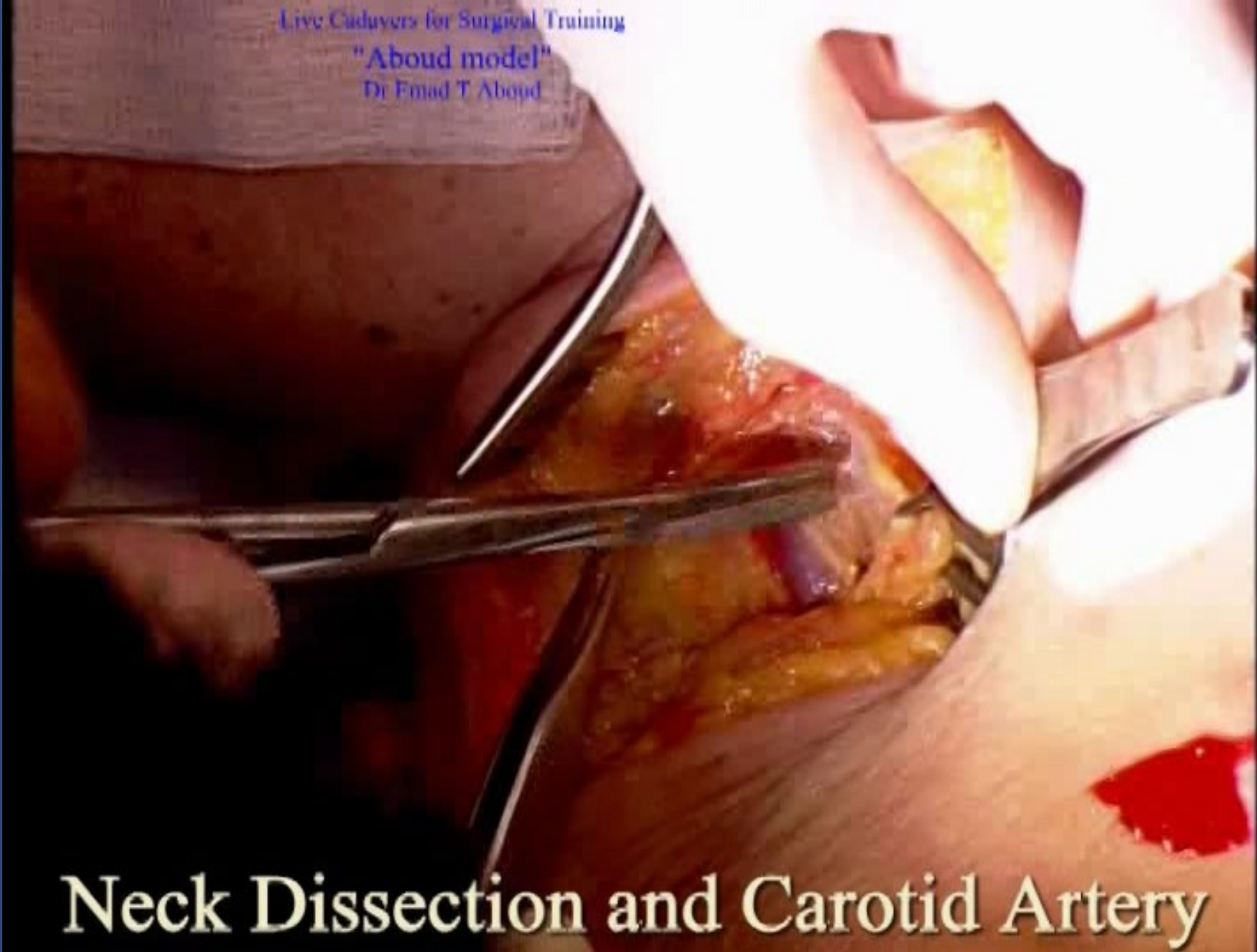




Live Cadavers for Surgical Training

"Aboud model"

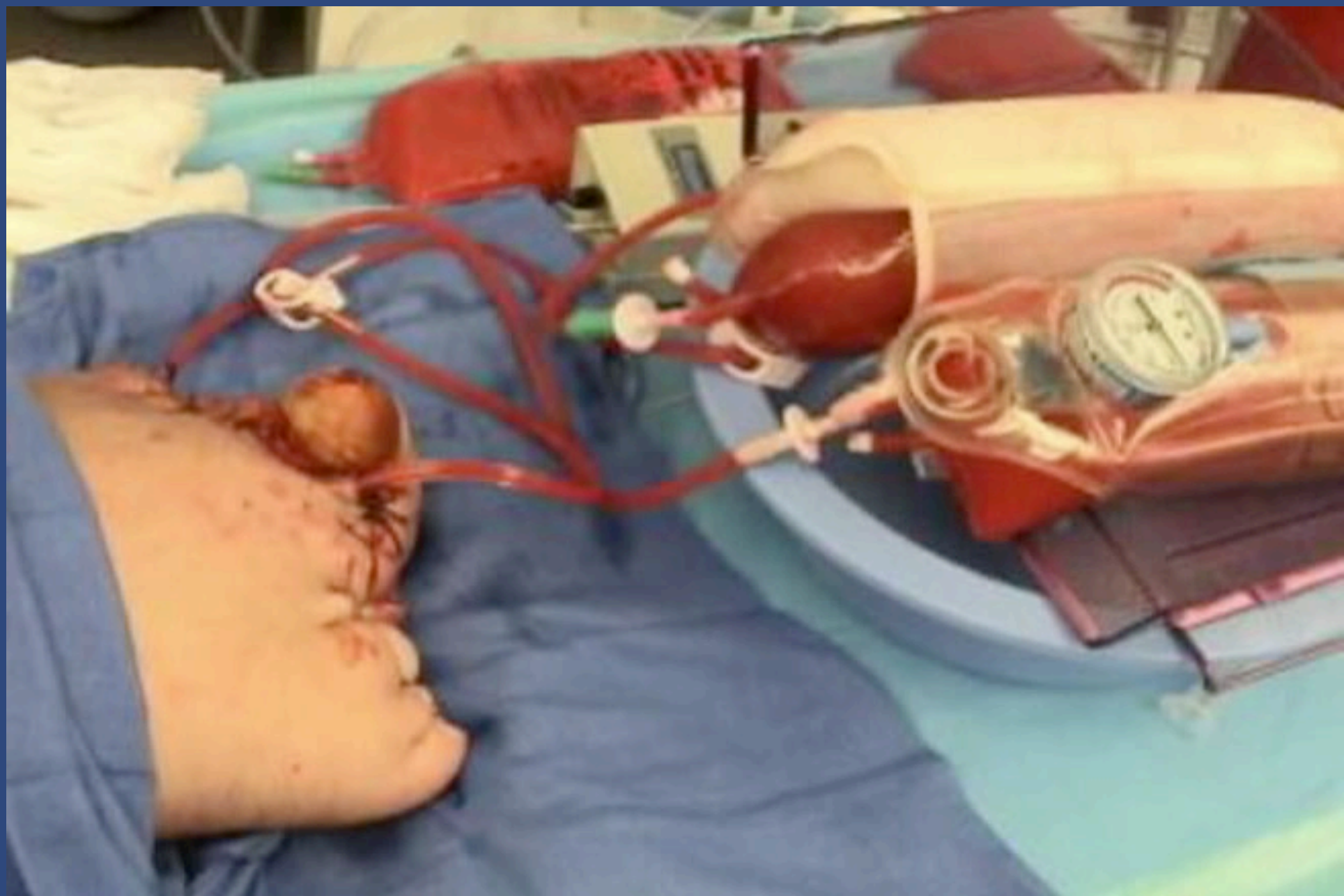
Dr Emad T Aboud



**Neck Dissection and Carotid Artery**











The cost of cadaver specimen is variable:

whole body	7000\$
Cephalous	1800\$
accessories: (Tubing and artificial blood) up to	500 \$
pulsatile pump or a secondhand heart pump	1500\$
Preparation, creating aneurysm models etc.	1500\$

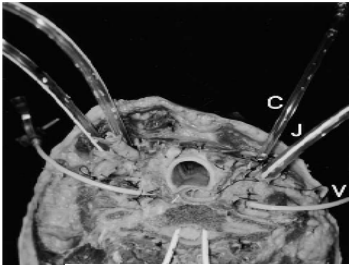


FIG. 1. ICA before craniotomy. C = inside the skull base; J = inside the skull base; V = vertebral artery.

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**Preparation**

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FIG. 2. Photograph demonstrating the connections between the cadaveric head and the reservoirs.

rate of 60 pulses per minute was selected; the machine provided a rate of 40 to 120 pulses/minute. Pressures up to 150 mm Hg can be applied through the pressure bag to the source of the red fluid. For our purposes, we applied a pressure of 80 mm Hg as a baseline be-

**Neurosurgical training model simulating live surgery**

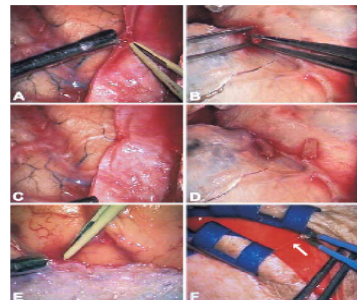
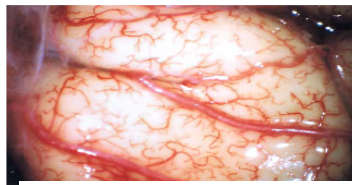


FIG. 3. Training session photographs showing opening and hemostasis. A and B: Bleeding meningeal arteries. C and D: The same arteries after coagulation. E: Coagulating pit vessels. F: Scalp hemostasis achieved using Raney clips and coagulation of vessels. Arrow indicates a bleeding jet.

agulated, or clamped using Raney clips. According to the intended procedure, a variety of craniotomies were performed, with care taken to preserve the underlying dura mater. The edges of the bone were waxed to prevent a fluid leak. The dura mater was opened and leaking vessels were coagulated (Fig. 3).

**Cerebral and Vascular Dissection.** The exposed brain was extremely lifelike (Fig. 4); the arteries were light red and pulsating, the veins were dark red and filled, and a clear fluid simulated the release of cerebrospinal fluid when the arachnoid was opened. We split the sylvian fissure and followed the branches of the MCA down to the carotid and basal cisterns, dissecting the branches of the circle of Willis and exposing all neurovascular structures in the skull base.

**Vascular Sparing and Anastomosis.** A variety of exercises were performed, starting with the establishment of an STA-MCA bypass (end-to-side anastomosis) and including repair of a longitudinal incision or a partial arterial defect and a transected artery (end-to-end anastomosis), as well as segmental arterial replacement. These procedures were performed on the cortical branches of the MCA and the M<sub>2</sub> and M<sub>3</sub> branches deep within the fissure. We used various segments of these branches. Each segment was dissected for approximately 1 cm of its length from the overlying arachnoid membrane. Small branches were coagulated and disconnected to free the segment. Two vascular clips were applied on both sides of the segment and arteriotomies were performed according to the kind of repair or anastomosis desired. After suture completion, the temporary clips were



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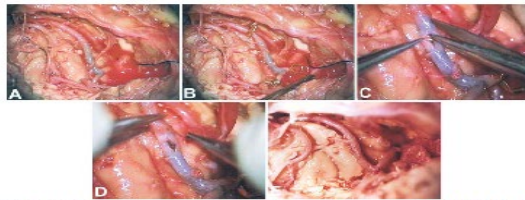


FIG. 6. Training session photographs showing thrombectomy and arterial repair of one of the M<sub>2</sub> branches. A: Thrombosis in the MCA branch. B: Temporary clipping. C: Arteriotomy. D: Removal of the thrombus. E: Establishment of flow through a patent vessel after arteriotomy.

**Resection of Artificial Tumors and Other Procedures.** Gelatinous material was injected into different locations of the basal cisterns and within the parenchyma to represent a tumor mass so that the trainee could practice resection of these masses while preserving neurovascular structures (Fig. 8). Skull base approaches, intracranial resections, and other procedures that we usually practice in cadavers prepared using traditional methods were practiced as well.

**Endoscopic Procedures.** After a frontal burr hole had been made, the sheath of the endoscope was introduced toward

the lateral ventricle. The optic apparatus was introduced after the introducer had been pulled out, and the choroid plexus and the septal and thalamic veins led the way to the foramen of Monro. The endoscope passed the foramen into the third ventricle and the mammillary bodies and the infundibular recess were identified. The floor of the third ventricle was perforated in front of the BA bifurcation in the area of the tuber cinereum. The BA trunk and branches, which were filled and pulsating, were identified in the interpeduncular cistern. Practicing irrigation and clearing of liquid inside the ventricles was achieved, as were observing the pul-

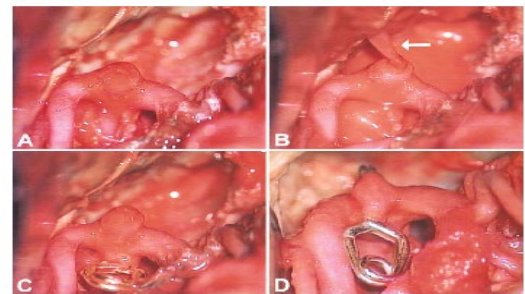


FIG. 7. Management of aneurysm bleeding. A: Artificial aneurysm located on the MCA bifurcation. B: Bleeding jet from the punctured aneurysm (arrow). C: Temporary clipping of the MCA. D: Clipping of the aneurysm neck.

# New laboratory model for neurosurgical training that simulates live surgery

EMAD ABOUD, M.D., OSSAMA AL-MEFTY, M.D., AND M. GAZI YAŞARGIL, M.D.

Department of Neurosurgery, University of Arkansas for Medical Sciences, Little Rock, Arkansas

**Object.** Laboratory training models are essential for developing and refining surgical skills, especially for microsurgery. The closer to live surgery the model is, the greater the benefit. In this paper the authors introduce a cadaver model with unique characteristics: dynamic filling of the cerebral vasculature with colored liquid and clear fluid filling of the arachnoid cisterns. This model is distinctive and has great practical value for training in a wide range of surgical procedures.

**Methods.** Cadaveric heads were prepared for surgical procedures in the following manner: the carotid arteries (CAs) and vertebral arteries (VAs) in the neck were cannulated, as were the internal jugular veins (IVs) on both sides. Two tubes were introduced into the spinal canal and each one was advanced into one of the cerebellopontine angle cisterns. A CA, VA, or both were then connected to a reservoir containing fast red fluid and a pressure of 80 to 120 mm Hg was maintained. A pulse rat connected to a vessels were clipped by an adjustable device and repaired; intracranial rupture; cavernoma; and aneurysm.

**Conclusions.** This model presents itself as an alternative to live surgery and refines surgical techniques.

**KEY WORDS**  
cerebral vas

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this paper: BA = basilar artery; MCA = middle cerebral artery; VA = vertebral artery

**Neurosurgical training model simulating live surgery**

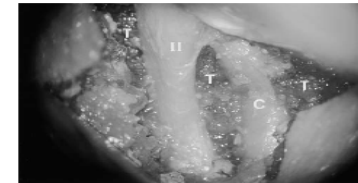


FIG. 8. Training session photograph obtained during the resection of an artificial parasellar tumor (T). C = carotid artery; II = optic nerve.

sation of the BA and identifying the liquid flow through the fenestra.

**Utilization of Both Hemispheres**

All the aforementioned procedures were then performed on the other side of the cadaveric head after the MCA on the first side had been closed and all ruptured vessels had been coagulated or clipped. The CA and the JV on the second side were connected to the colored fluid reservoirs. (Changing connections from side to side between the VA and the JV and the fluid reservoirs had no impact.) The same procedures were then performed on the second side in addition to interhemispheric approaches.

**Posterior Circulation**

When all possible training procedures on the anterior circulation had been completed, the CA was disconnected and the posterior communicating arteries were clipped on both sides proximal to the posterior cerebral arteries. The VA from one or both sides was connected to the reservoir filled with light red fluid and one of the IVs was connected to the source of the dark red fluid. Dissection of the posterior fossa and posterior circulation, in addition to previously mentioned procedures, were then performed through the occipital and suboccipital approaches.

**Whole-Brain Application**

We also applied this method in a whole-brain specimen obtained at autopsy. In this case both CAs were cannulated, allowing a variety of vascular exercises on the major branches (Fig. 9).

**Discussion**

As a surgical specialty, neurosurgery requires the development of dexterity and skills for basic and challenging procedures and techniques. In delicate organs such as the central nervous system, the surgeon's individual skills play a crucial role in determining patient outcome. Hence, the emphasis has been placed on laboratory training, preparing surgical trainees for the operating room experience.<sup>10</sup> The



FIG. 9. Photograph demonstrating cannulation of the CA in a brain obtained at autopsy.

fine manipulation and dissection of cerebral vessels with anastomosis usually have been practiced on animals.<sup>11</sup> Unfortunately, these procedures are limited to a simple technique and have no relation to the actual anatomy or to surgical crises that are encountered by the trainee during live surgery. A critical part of this training is mastering the anatomy. Hence, fine publications, methods, and courses are richly introduced and widely available.

To improve the illustrative value of cadaver dissection, colored materials are injected into the vessels of cadavers to identify arteries and veins for anatomical studies. Fluorescein and radiopaque substances, silicone, gelatin, latex, acrylic, or tinted polyester resin<sup>12,13</sup> have been used for this purpose. Mechanical pressure pumps have been used to introduce and perfuse embalming fluids via the common CAs or femoral arteries.<sup>14</sup> Nevertheless, there have been no reports of using such machines to induce pulsation and vascular filling in cadavers for training purposes. In studying the role of neurovascular compression in trigeminal neuralgia, Harnlyn<sup>15</sup> described injection filling of cadaveric vessels to determine neurovascular relationships within the posterior fossa.

To our knowledge, a model such as ours has not previously been developed. This model can increase the capacity of neurosurgical laboratories to train for a variety of surgical approaches, including skull base, neurovascular, endoscopic, and even endovascular procedures. The presence of clear fluid in the subarachnoid spaces, the pulsation, and the vascular filling give greater realism to these training procedures. This model provides the trainee with a unique

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## “Live cadavers” for training in the management of intraoperative aneurysmal rupture

Emad Aboud, MD,<sup>1</sup> Ghaith Aboud,<sup>2</sup> Ossama Al-Mefty, MD,<sup>3</sup> Talal Aboud,<sup>2</sup> Stylianos Rammos, MD,<sup>1</sup> Mohommad Abolfotoh, MD, PhD,<sup>3,4</sup> Sanford P. C. Hsu, MD,<sup>5</sup> Sebastian Koga, MD,<sup>6</sup> Adam Arthur, MD, MPH,<sup>7</sup> and Ali Krisht, MD<sup>1</sup>

<sup>1</sup>Arkansas Neuroscience Institute, St. Vincent Health System, Little Rock, Arkansas; <sup>2</sup>Atlantic University, School of Medicine, Island Park, New York; <sup>3</sup>Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts; <sup>4</sup>Department of Neurosurgery, Ain Sham University, Cairo, Egypt; <sup>5</sup>Taipei Veterans General Hospital, National Yang-Ming University, Taipei, Taiwan; <sup>6</sup>International Neuroscience Institute, Hannover, Germany; and <sup>7</sup>Semmes-Murphey Neurologic and Spine Institute, and Department of Neurosurgery, University of Tennessee, Memphis, Tennessee

**OBJECT** Intraoperative rupture occurs in approximately 9.2% of all cranial aneurysm surgeries. This event is not merely a surgical complication, it is also a real surgical crisis that requires swift and decisive action. Neurosurgical residents may have little exposure to this event, but they may face it in their practice. Laboratory training would be invaluable for developing competency in addressing this crisis. In this study, the authors present the “live cadaver” model, which allows repetitive training under lifelike conditions for residents and other trainees to practice managing this crisis.

**METHODS** The authors have used the live cadaver model in 13 training courses from 2009 to 2014 to train residents and neurosurgeons in the management of intraoperative aneurysmal rupture. Twenty-three cadaveric head specimens harboring 57 artificial and 2 real aneurysms were used in these courses. Specimens were specially prepared for this technique and connected to a pump that sent artificial blood into the vessels. This setting created a lifelike situation in the cadaver that simulates live surgery in terms of bleeding, pulsation, and softness of tissue.

**RESULTS** A total of 203 neurosurgical residents and 89 neurosurgeons and faculty members have practiced and experienced the live cadaver model. Clipping of the aneurysm and management of an intraoperative rupture was first demonstrated by an instructor. Then, trainees worked for 20- to 30-minute sessions each, during which they practiced clipping and reconstruction techniques and managed intraoperative ruptures. Ninety-one of the participants (27 faculty members and 64 participants) completed a questionnaire to rate their personal experience with the model. Most either agreed or strongly agreed that the model was a valid simulation of the conditions of live surgery on cerebral aneurysms and represents a realistic simulation of aneurysmal clipping and intraoperative rupture. Actual performance improvement with this model will require detailed measurement for validating its effectiveness. The model lends itself to evaluation using precise performance measurements.

**CONCLUSIONS** The live cadaver model presents a useful simulation of the conditions of live surgery for clipping cerebral aneurysms and managing intraoperative rupture. This model provides a means of practice and promotes team management of intraoperative cerebrovascular critical events. Precise metric measurement for evaluation of training performance improvement can be applied.

<http://thejns.org/doi/abs/10.3171/2014.12.JNS141551>

**KEY WORDS** live cadaver; perfused cadaver; surgical training; cerebral aneurysm; pulsatile model; neurovascular

**T**HE intraoperative rupture of an intracranial aneurysm can be a critical event that can jeopardize a patient's outcome. The rate of this event ranges from 6% to 20% of all surgical cases.<sup>5,9,11,14,16</sup> In some reports, the aneurysmal rupture rate is as high as 41.6%.<sup>21</sup> In

a recent study that reviewed the literature, involving a total of 9488 aneurysm surgeries, Madhugiri and colleagues<sup>17</sup> found that the rate of intraoperative rupture in the entire group was 9.2%.

Factors such as the reduced number of working hours

**ABBREVIATION** MCA = middle cerebral artery.

**SUBMITTED** July 11, 2014. **ACCEPTED** December 11, 2014.

**INCLUDE WHEN CITING** Published online July 3, 2015; DOI: 10.3171/2014.12.JNS141551.

**DISCLOSURE** The technique described in this article is based on US Patent no. 6,790,043 (2004): Method and apparatus for surgical training. This work was sponsored in part by the institutions mentioned in the body of the article: Arkansas Neurosciences Institute, Little Rock, Arkansas; Practical Anatomy Workshop, St. Louis, Missouri; Medical Education and Research Institute, Memphis, Tennessee; and National Yang-Ming University, Taipei, Taiwan.

## “Live Cadavers” for Practicing Airway Management

Emad T. Aboud, MD; Ghaith Aboud; Talal Aboud

**ABSTRACT** Human cadavers have been used successfully as training models to practice airway management, but the lack of lifelike conditions reduces the utility of this model when softness of tissue and the ability to bleed are required for training scenarios. This report describes our “live cadaver” model, which combines lifelike conditions with real human anatomy. Five human cadavers were prepared as “live cadavers”. This entailed cannulating the carotid and femoral arteries and the jugular and femoral veins, and then connected them to artificial blood reservoirs. An intra-aortic balloon pump was used to provide pulsating flow through the heart and major arteries. Finally, central and peripheral lines were inserted. Multiple techniques related to airway management were practiced in setting simulating the treatment of casualties with multiple trauma to include emergency cricothyroidotomy. With this model, participants were confronted with medical situations similar to those found in traumatized live patients (e.g., blood and other body fluids filling the mouth and nose, edema of the tongue and face). With the combination of lifelike conditions and real human anatomy, our experience demonstrated that the “live cadaver” increased the training value of traditionally prepared cadaver models.

### INTRODUCTION

Laryngoscopy, intubation, and other airway management techniques are commonly taught in elective anesthesia settings on live patients.<sup>1</sup> Although this situation provides the ideal setting, it creates problems that severely limit physician training.<sup>2</sup> Patient safety is the priority, and procedures cannot be repeated just for practice. As an alternative, cadavers offer a unique opportunity for airway management training because the physician can interact with, and visualize, human tissue without the clinical risks associated with practicing on live patients. In addition to cadavers, live animals, animal heads, mannequins, and virtual reality have all been used for this purpose.<sup>3-7</sup>

Most of these models are extremely important in teaching the individual skills required for airway management. However, an important feature missing in these models is the combination of the lifelike conditions of the living body with real human anatomy. Such a combination is essential for advanced training. Some reports suggest that adjunctive teaching sessions that use cadavers do not improve the physician’s initial success in clinical intubation over the use of the mannequin alone, especially for repetitive training.<sup>8,9</sup> We believe that adding the lifelike conditions to cadaver models (“live cadaver”) will overcome some of the shortcomings encountered with available training models.<sup>10-15</sup> In addition, the “live cadaver” model enhances skills acquisition for airway management, and promotes teamwork.

This model was not initially developed for training on airway management techniques, but the success and positive

feedback we achieved in using this model for neurosurgical, vascular, and trauma surgery<sup>16-18</sup> encouraged us to use it for other surgical and procedural techniques as an advanced level of physician training. This model was also used to test medical devices with clinical applications<sup>19</sup> and in conjunction with other training techniques.<sup>20,21</sup> In this report, we describe the use of a technique that combines the lifelike conditions of the living body with real human anatomy for training on airway management procedures.

### METHODS

All cadavers were ethically donated according to the individual’s legally executed, advance directive bequest on file at the host site. Sponsoring institutions included the University of Arkansas for Medical Sciences, Little Rock, Arkansas; Upstate Medical University, Syracuse, New York; Saint Louis University, Saint Louis, Missouri; and the Anatomy Gift Registry, Glen Burnie, Maryland. The “live cadaver” model, which was developed initially for neurosurgery training, is in practical use in some of these centers and in other academic and nonacademic training centers for surgical training in different surgical disciplines.

### Preparation

Five fresh, whole human cadavers were cannulated and prepared according to the preparation protocol for the “live cadaver” model. Each of the five cadavers was placed supine on an embalming gurney and washed with an antibacterial soap. The Duotronic embalming machine (California Professional Manufacturing, Modesto, California) was filled with saline to prepare for the infusion. A small incision was made along the lower anterior borders of the sternocleidomastoid muscle, and the right internal jugular vein and common carotid artery were exposed. Ligatures were looped around the jugular and the carotid artery, and a small incision was then made on the jugular vein. Long forceps were inserted

## Novel Simulation for Training Trauma Surgeons

Emad T. Aboud, MD, Ali F. Krisht, MD, Terence O’Keeffe, MD, Remi Nader, MD, Moustafa Hassan, MD, C. Melinda Stevens, MD, Fahd Ali, MD, and Fred A. Luchette, MD, MSc

**Background:** Clinical training in operative technique is important to boost self-confidence in residents in all surgical fields but particularly in trauma surgery. The fully trained trauma surgeon must be able to provide operative intervention for any injury encountered in practice. In this report, we describe a novel training model using a human cadaver in which circulation in the major vessels can be simulated to mimic traumatic injuries seen in clinical practice.

**Methods:** Fourteen human cadavers were used for simulating various life-threatening traumatic injuries. The carotid and femoral arteries and the jugular and femoral vein were cannulated and connected to perfusate reservoirs. The arterial reservoir was connected to an intra-aortic balloon pump, which adds pulsatile flow through the heart and major arteries.

Fully trained trauma surgeons evaluated the utility of this model for repairing various injuries in the thoracic and abdominal cavity involving the heart, lungs, liver, and major vessels while maintaining emergent airway control.

**Results:** Surgeons reported that this perfused cadaver model allowed simulation of the critical challenges faced during operative trauma while familiarizing the student with the operative techniques and skills necessary to gain access and control of hemorrhage associated with major vascular injuries.

**Conclusion:** In this report, we describe a novel training model that simulates the life-threatening injuries that confront trauma surgeons. An alternative to living laboratory animals, this inexpensive and readily available model offers good educational value for the acquisition and refinement of surgical skills that are specific to trauma surgery.

**Key Words:** Surgical training, Trauma training, Vascular, Training model, Simulation, Alternative model, Surgical skills, Live surgery, Live cadaver, Perfused cadaver, Stab wounds, Military surgical training, Combat injury, Circulation, Bleeding, Cadaver, Anatomy, Veterinary surgical training, Education.

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Supported by the University of Arkansas for Medical Sciences, Little Rock, Arkansas; Arkansas Neurosciences Institute, Little Rock, Arkansas; People for Ethical Treatment of Animals, USA, State University of New York, Syracuse, New York; Practical Anatomy Workshop Saint Louis University, Saint Louis, Missouri.

Supplemental digital content is available for this article. Direct URL citations appear in the printed text, and links to the digital files are provided in the HTML text of this article on the journal’s Web site ([www.jtrauma.com](http://www.jtrauma.com)).

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DOI: 10.1097/TA.0b013e3182396337

Arkansas Neuroscience Institute, Saint Vincent Infirmary, 5 Saint Vincent Circle, Little Rock, AR 72205.

This article was presented at the Military Health System Research Symposium, Fort Lauderdale, Florida, August 12-15, 2013.

This work describes a method patented by the first author: U.S. Patent number 6790043.

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US006790043B2

(12) **United States Patent**  
**Aboud**

(10) **Patent No.:** **US 6,790,043 B2**  
(45) **Date of Patent:** **Sep. 14, 2004**

(54) **METHOD AND APPARATUS FOR SURGICAL TRAINING**

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(List continued on next page.)

(75) Inventor: **Emad T. Aboud**, Swaida (SY)

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(73) Assignee: **Board of Trustees of the University of Arkansas**, Little Rock, AR (US)

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(List continued on next page.)

(\* ) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 57 days.

*Primary Examiner*—Derris H. Banks  
*Assistant Examiner*—Dmitry Suhol

(21) Appl. No. 10/220,052

The model is recognized and sited in the Medical Modeling and Simulation data base of the American College of Surgeon



**AMERICAN COLLEGE  
OF SURGEONS**

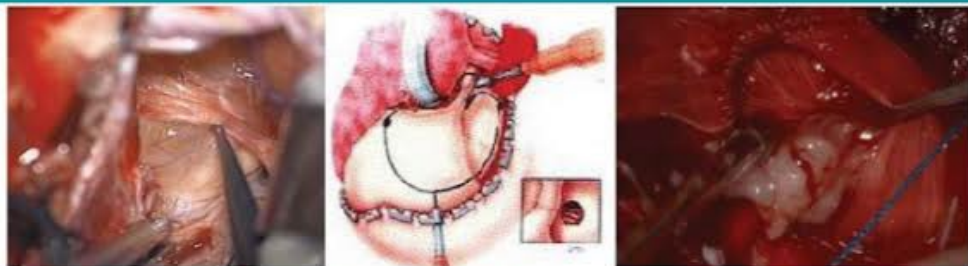
## Medical Modeling and Simulation Database

#	Edit	Rank	Title	Date of Publication	Full Author	Publication Type
1.		77	<a href="#">New laboratory model for neurosurgical training that simulates live surgery.</a>	2002 Dec	<b>Aboud, Enad; Al Mefly, Ossama; Yasargil, M Gazi</b>	Journal Article
2.		62	<a href="#">Robotic radical prostatectomy: a technique to reduce pT2 positive margins.</a>	2004 Dec	Ahlering, Thomas E; Eichel, Louis; Edwards, Robert A; Lee, David I; Skarecky, Douglas W	Journal Article









## Al-Mefty Residents Course, Microsurgery of the Skull Base

Hands-on Cadaver Workshop with Live Surgery & Life-Like Simulation

Sep 20-22, 2025

### Ossama Al-Mefty, MD

Director of Skull Base Program  
Harvard School of Medicine - Boston, MA

#### Invited Faculty

##### **Kaith Almefty, MD**

Barrow Neurological Institute, Phoenix, AZ

##### **Kenan Arnautovic, MD**

University of Tennessee, Memphis, TN

##### **Luis A Borba, MD**

Federal University of Parana, Curitiba, Brazil

##### **Mark Eisenberg, MD**

Cushing Neuroscience Institute, Great Neck, NY

##### **Marcio Rassi, MD**

Evangelic University, Curitiba, Brazil

##### **Paulo A.S. Kadri, MD**

Federal University, Campo Grande, Brazil

##### **Rami Almefty, MD**

Lewis Katz School of Medicine, Philadelphia, PA

##### **Samer Ayoubi, MD**

Abbaseen Hospital, Damascus, Syria

##### **Walid Ibn Essayed, MD**

McGovern Medical School, UTHealth, Houston, TX

##### **Wenya Linda Bi, MD**

Harvard School of Medicine, Boston, MA

### Course Director

#### Ali Krisht, MD

Director, Arkansas Neuroscience Institute

#### ANI Faculty

##### **Emad Aboud, MD**

##### **Stylios Rammos, MD**

##### **Tarek Abuelem, MD**

##### **Khaled Krisht, MD**

##### **Lucas Possatti, MD**

##### **Mateus Neto, MD**

##### **Jaafar Basma, MD**

##### **Paul Lee, MD**

##### **Li Cai, MD**

##### **Hossam Youssef, MD**

##### **Abdel Raouf Kayssi, MD**



## M.G. YASARGIL ADVANCED COURSE IN CEREBRAL ANEURYSMS

Hands-On Cadaver Workshop with Live Surgery & Life-Like Simulation

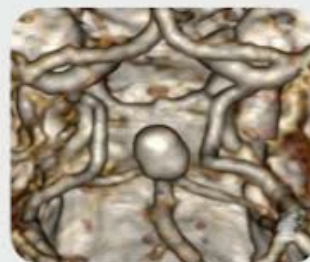
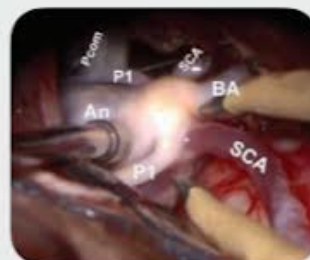


### COURSE DIRECTOR:

Ali Krisht, M.D.

### ANI FACULTY:

Emad Aboud, M.D.	Li Cai, M.D.
Stylios Rammos, M.D.	Paul Lee, M.D.
Tarek Abuelem, M.D.	Hossam Youssef, M.D.
Khaled Krisht, M.D.	Maher Hulou, M.D.
Jaafar Basma, M.D.	Abdel Raouf Kayssi, M.D.



### GUEST FACULTY:

Ketan Ramanlal Bulsara, M.D.

University of Connecticut Farmington, Connecticut

Kathleen Joos, M.D.

St. Dimpna Hospital, Ziekenhuis Oost-Limburg Genk

Jean de Oliveira, M.D.

Santa de Casa de São Paulo, Brazil

Lassaad Bsili, M.D.

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Lucas Possatti, M.D.

Bela Vista, São Paulo, Brazil

Mateus Reghin Neto, M.D.

Bela Vista, São Paulo, Brazil

Miguel Lopez-Gonzalez, M.D.

Loma Linda University, California

Sanford P.C. Hsu, M.D.

National Yang-Ming University, Taipei, Taiwan

Wei-Hsun Yang, M.D.

New Taipei Municipal TuCheng Hospital, Taipei, Taiwan



## Al-Mefty Residents Course, Microsurgery

Hands-on Cadaver Workshop with Live Surgery

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Director of Skull Base Program  
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McGovern Medical School, UTHealth, Houston, TX

**Wenya Linda Bi, MD**

Harvard School of Medicine, Boston, MA

# Arkansas Neuroscience Institute

Hands-on Cadaver Workshops, Live Surgery & Life-Like Simulation

## ANI Live Microneurosurgery Course

March 9-13, 2025



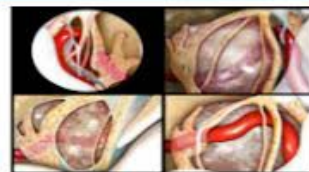
## M.G. Yasargil Advanced Course in Cerebral Aneurysms

May 3 - 5, 2025



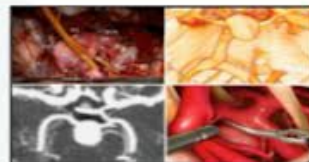
## Evandro de Oliveira Cavernous Sinus Course

June 21 - 24, 2025



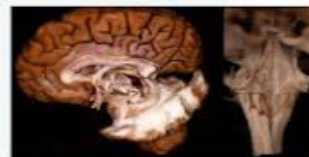
## Al-Mefty's Course, Microneurosurgery of the Skull Base

Sept 20 - 22, 2025



## Advanced Course in Fiber Dissection

Nov 15-17, 2025



**O. Al-Mefty Microneurosurgery Laboratory** is a state-of-the-art microsurgical laboratory with fully equipped working stations that also has the "Live Cadaver" model that circulates a blood-like perfusate throughout the vessels for neurosurgical training

## ADVANCED COURSE NEURYSMS

Surgery & Life-Like Simulation



### COURSE DIRECTOR:

Ali Krisht, M.D.

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New Taipei Municipal TuCheng Hospital, Taipei, Taiwan

# Finally

The Live Cadaver Model  
is the only  
Existed model that combines  
the real human anatomy  
with the life-like conditions  
of the living body

[aboudemad@gmail.com](mailto:aboudemad@gmail.com)

**Thank you**